

Right Methods to Extend the Meat Shelf- Life by Using of Natural Preservatives and Their Public Health Importance

Fahim A. Shaltout^{1*}

¹Food Control, Faculty of Veterinary Medicine, Benha University, Egypt.

Received date: 02 September 2024; **Accepted date:** 16 September 2024; **Published date:** 21 September 2024

Corresponding Author: Fahim A. Shaltout, Food Control, Faculty of Veterinary Medicine, Benha University, Egypt. <https://orcid.org/0000-0002-8969-2677>

Citation: Fahim A. Shaltout. Right Methods to Extend the Meat Shelf- Life by Using of Natural Preservatives and Their Public Health Importance. Journal of Medicine Care and Health Review 1(2). <https://doi.org/10.61615/JMCHR/2024/SEPT027140921>

Copyright: © 2024 Fahim A. Shaltout. This is an open-access article distributed under the terms of the Creative Commons Attribution License, which permits unrestricted use, distribution, and reproduction in any medium, provided the original author and source are credited.

Abstract

Meat is an excellent source of nutrients for human beings. Meat provides a favorable environment for microbial growth. Prevention of microbiological contamination of meat, chemical preservatives, including nitrites, nitrates, and sorbates, have been used in food processing due to their low cost and strong antibacterial activity. The application of chemical preservatives is recently being considered by customers due to concerns related to negative population health issues. Demand for natural substances as food preservatives has increased with the use of plant-origin and animal-origin foods and microbial metabolites. In regarding Natural preservatives inhibit the growth of spoilage bacteria or foodborne pathogens by increasing the permeability of microbial cell membranes, interrupting protein synthesis, and cell metabolism. By using Natural preservatives, we can extend shelf-life and inhibit the growth of bacteria. In regarding to natural preservatives can affect food sensory properties, including flavor, taste, color, texture, and acceptability of food. In regard to Increasing the applicability of natural preservatives, a number of strategies, including combinations of different preservatives or food preservation methods, such as active packaging systems and encapsulation, have been used for applications of methods of natural preservatives in meat.

Key Words: Meat, preservation methods, packaging, food-borne pathogens, population health.

Introduction

Food-borne pathogens, including *Listeria monocytogenes*, *Staphylococcus aureus*, pathogenic *Escherichia coli*, *Clostridium perfringens*, *Campylobacter* spp., and *Vibrio* spp., cause a large number of illnesses, with substantial damage to population health and economy. World Health Organization, food contaminated with food-borne pathogens, chemicals, and allergens results in 600 million cases of foodborne illness and four hundred thousand deaths worldwide/ year, Fifty-six million people die /year and 7.7% of people worldwide suffer from food-borne diseases. Meat is an important nutrient source for human beings due to its excellent protein content, essential amino acids, vitamin B groups, and minerals. Meat provides an appropriate environment for spoilage bacteria or food-borne pathogens due to its high water activity and important nutrient factors [1,2,3,4,5,6,7]. Food processing has advanced worldwide, resulting in an enhanced threat of food contamination by food-borne pathogens, chemical residues, harmful food additives, and toxins. Multiplication of spoilage and pathogenic bacteria should be controlled to ensure food safety. Food preservation methods for protecting food from food-borne pathogens and extending shelf-life include chemical methods, such as the use of preservatives; physical methods, such as heat treatment, drying, freezing, and packaging; and biological methods using bacteria that have an antagonistic effect on pathogenic bacteria and produce bacteriocins. The addition of food preservatives that inhibit the growth of bacteria is a widely used food protection technique. Countries in the world have different regulations for food preservatives [8,9,10,11,12,13,14]. Chemical preservatives have an advantage for meat production due to low cost, guaranteed antibacterial effect or shelf-life extending activity, and little action on the taste, flavor, color, and texture of

meat. Chemical means of meat preservatives tend to be less preferred by consumers because of a number of population health concerns regarding their side actions. Consumers selected preservatives as the most concerned food additive owing to their negative action on population health. Sorbic acid, benzoic acid, and their salts promote mutagenic and carcinogenic compounds. Both methods, Nitrites, and nitrate, used as preservative and coloring agents in meat preservation methods, were associated with leukemia, colon cancer, bladder cancer, and others. Methods of Natural preservatives have emerged as alternatives to chemical preservatives. Methods of Natural preservatives have shown potential to provide effective antimicrobial activity and reduce negative population health action. Meat contaminated by chemical additives is a major concern for population health. Meat producers and researchers have begun to consider the use of natural rather than chemical preservatives [15,16,17,18,19,20,21]. ‘Clean label’ food trends, including meat and its products, began and possessed an important source of food marketing. It includes consumer-friendly characteristics, such as chemical additive-free, least processing, a brief list of food ingredients, and the procedure of traditional methods. clean label food material market, including natural preservatives, is likely to value, mostly owing to growing consumer requests for all-natural products. Natural preservatives such as nisin, natamycin, ϵ -polylysine, and grapefruit seed extract are registered, but they are not approved for meat, or their concentration is not specified. Replacement of chemical preservatives with natural preservatives has major positive action and is being accepted by customers. Food producers also encounter challenges, including a decrease in price competitiveness due to the relatively high price of natural preservatives and a decrease in

antibacterial effect due to food ingredients, such as carbohydrates, proteins, and lipids. In the case of plant-origin substances, standardization is problematic because of the influence of country of origin, soil, and harvest seasons. Toxicity evaluation or identification of exact compounds for several plant-origin compounds contained in extracts and essential oils have been performed. Solving these problems, various studies have been conducted to optimize the extraction process, combine other antimicrobial substances, apply active packaging, and encapsulate antibacterial substances to improve their utilization [22,23,24,25,26,27,28]. This review summarizes current knowledge about the application of natural preservatives for meat against food-borne pathogens and spoilage bacteria.

Application Technique of Natural Preservatives to Meat and its Products

Natural preservatives are manufactured in a variety of formulations including powder formed by drying methods and liquid forms such as essential oils. Natural preservatives are directly added to meat and extend shelf-life by inhibiting bacterial growth. It is possible to increase the antibacterial effect of natural preservatives through a combination of other food processing methods [29,30,31,32,33,34,35]. In the case of plant-origin natural preservatives, it is necessary to consider the form applied to food. Natural preservatives are commonly prepared in the form of extracts using organic solvents, water, and essential oils. plant extracts obtained from rosemary, chestnut, sage, cranberry, oregano, grape seed, and others have been used as meat preservatives. application of plant-origin substances to meat products in the form of essential oil because the antibacterial effect of the essential oil type is better than that of the extract type. It is difficult to apply large amounts of essential oil to food because of its distinct organoleptic properties. Recent developments have attempted to solve this problem by applying essential oils with other antibacterial substances. advantage of this application is that it reduces amounts of essential oils with strong flavors and increases antioxidant and antibacterial action through synergistic action. In terms of an industrial perspective, if chemical preservatives cannot be completely replaced with natural preservatives, due to industrial problems, such as increasing economic costs or complexity of the product manufacturing process, they could be replaced gradually by composing a mixed formulation of chemical preservatives and natural preservatives [36,37,38,39,40,41,42]. gamma irradiation and high-pressure processing (HPP) treatment are physical food-processing methods that can further increase the antibacterial efficacy of natural preservatives. Unlike thermal food processing, these two food processing techniques could be used for the pasteurization of raw meat because it has a minor effect on food composition. In 1997, WHO, the Food and Agricultural Organization (FAO), and the International Atomic Energy Agency (IAEA) concluded that foods processed in proper doses of irradiation are nutritionally sufficient and safe to consume. Irradiation is permitted for food preservation methods in more than sixty countries. Recent approaches in food irradiation have involved the use of combined treatments with natural preservatives to reduce irradiation doses. Gamma irradiation of medium doses (2–6 kGy) with natural compounds and active packaging has been applied to extend the shelf-life of meat. HPP is also a non-thermal method for food preservation that inhibits the growth of bacteria and maintains food's natural properties. HPP is performed under high pressures (100–800 MPa) at

mild temperatures or weak heating. The potential capability of combining HPP and natural preservative methods including essential oil and antibacterial peptides in alleviating both processing conditions of HPP and concentration of natural preservatives while maintaining antibacterial action [43,44,45,46,47,48,49]. Encapsulation is one of the effective methods for expanding the applicability of natural preservatives to food. Encapsulation was performed with GRAS (generally recognized as safe) materials such as alginate, chitosan, starch, dextrin, and proteins using various techniques including spray drying, extrusion, freeze drying, coacervation, and emulsification. The use of natural preservatives in meat is limited due to their characteristics, such as low solubility and bioavailability, rapid release, and easy degradation. Environmental conditions, such as pH, storage temperature and time, oxygen, and light exposures could influence the efficacy of natural preservatives. Through encapsulation, natural preservatives, especially hydrophobic compounds (e.g., essential oil), could improve its stability and expand the versatility of food processing while maintaining an antibacterial effect [50,51,52,53,54,55,56]. Active packaging is an innovative packaging technology that allows for an interaction with a product and its environment to extend shelf- -life and ensure its microbial safety while keeping the original properties of packaged food. In relation to European Union Guidance to Commission Regulation, active packaging is a type of food packaging with a further beneficial function, while providing a protective barrier against external influence. In meat processing, antimicrobial active packaging could be applied in several methods which are the incorporation of natural preservatives into a sachet inside packaging, packaging film composition with natural preservatives, packaging coated with natural preservatives onto the surface of food, and use of antimicrobial polymers as packaging materials [57,58,59,60,61,62,63]. Application of microorganism origin natural preservatives, known as biopreservation methods, in which useful bacteria or their antibacterial substances have antagonistic action on pathogenic or spoilage bacteria, are also a meat preservation method in the spotlight. The method is mainly applied in lactic acid bacteria, *Lactobacillus* spp., *Leuconostoc* spp., *Pediococcus* spp., and *Lactococcus* spp., that have a GRAS status, widely participate in fermentation processes, and produce various antibacterial metabolites such as organic acids, hydrogen peroxide, and bacteriocins methods. In terms of application to meat products, biopreservation methods included direct inoculation with lactic acid bacteria, which has an inhibitory effect on spoilage or pathogenic bacteria, the inclusion of bacterial strains producing antimicrobial substances in fermentation starter, and treatment with purified bacteriocins [64,65,66,67,68,69,70].

Natural Preservatives from Plants and Their Application for Meat and its Products

The antibacterial effect of plant-origin natural preservatives is closely related to polyphenols, phenolics, and flavonoids. Plant-origin polyphenols have various classifications and structures, as phenolic acids (caffeic acid, rosmarinic acid, gallic acid, ellagic acid, cinnamic acid), flavones (luteolin, apigenin, chrysoeriol), flavanols (catechin, epicatechin, epigallocatechin, gallic acid, and their gallate derivatives), flavanones (hesperidin, hesperetin, heridictyol, naringenin), flavonols (quercetin, kaempferol,

myricetin), isoflavones (geinstein, daidzin, formononetin), coumarins (coumarin, warfarin, 7-hydroxycoumarin), anthocyanins (pelargonidin, delphinidin, cyanidin, malvidin), quinones (naphthoquinones, hypericin), alkaloids (caffeine, berberine, harmaline), and terpenoids (menthol, thymol, lycopene, capsaicin, linalool) [71,72,73,74,75,76,77]. Polyphenols have been recognized for their effective antimicrobial properties. Although the antimicrobial mechanism has not yet been clearly elucidated, cell membranes disturb molecules, such as hydroxy group (OH-), which induces leakage of intracellular components, inactivation of metabolic enzymes, and extinction of adenosine triphosphate (ATP) structure; direct pH change in environment by improvement in proton concentration, reduction of intracellular pH by separation of acid molecules, and modification of bacterial membrane permeability; an organic acid in plant extracts may influence the oxidation of nicotinamide adenine dinucleotide (NADH), eliminating, reducing agent used in electron transport system [78,79,80,81,82,83,84].

Rosemary

Rosemary (*Rosmarinus officinalis* L) is a perennial herb with woody, aromatic, and evergreen needle-like leaves. Originally from the Mediterranean region, it is broadly distributed throughout the globe. Rosemary has been used in food as a spice and flavoring agent. Rosemary essential oil is known to contain fifteen kinds of bioactive compounds. The principal compound was 1,8-cineole (35.32%). Other major compounds were camphor, α -pinene, trans-caryophyllene, α -thujone, and borneol [85,86,87,88,89,90,91]. Antibacterial effect of rosemary ethanol extracts against *Listeria monocytogenes* in beef. Application of 45% rosemary ethanol extract for *Listeria monocytogenes* on beef led to a two-log CFU / gram reduction in incubation at 4 °C for nine days. In chicken meat, the effect of rosemary essential oil on inhibition of *Salmonella* Enteritidis and spoilage protective action at four and 18 °C was investigated. Five mg/mL of rosemary essential oil induced a decrease in coliform, aerobic bacteria, lactic acid bacteria, and anaerobic bacteria at 18 °C for one day. Compared with untreated chicken meat, reductions of 1.75 log CFU / gram (coliform), 0.87 log CFU / gram (aerobic bacteria), 1.05 log CFU / gram (lactic acid bacteria), and 1.28 log CFU / gram (anaerobic bacteria) were observed in the group treated with rosemary essential oil at 18 °C. Rosemary oil reduced *S. Enteritidis* by more than two log Colony Forming Units/gram at 18 °C, but less than one log CFU / gram at 4°C [92,93,94,95,96,97,98]. Rosemary essential oil applied with modified atmosphere packaging for inhibition of food-borne pathogens such as *S. Typhimurium* and *Listeria monocytogenes* in poultry filets under refrigerated conditions for seven days was examined. The 0.2% rosemary essential oil did not affect the sensory profile and inhibited the growth of food-borne pathogens in laboratory media within one day. Treatment with 0.2% rosemary essential oil did not affect the reduction in *S. Typhimurium* but showed weak antibacterial activity against *Listeria monocytogenes* until the first day of storage (0.1 log CFU / gram reduction compared to control) [99,100,101,102,103,104,105].

Sage

Sage (*Salvia officinalis* L.), belonging to the Lamiaceae family, has been used since the prehistoric eras because of its flavor, taste, therapeutic, and preservative properties. Sage is known to contain considerable amounts of

rosemary acid, p-coumaric acid, and benzoic acid. Sage essential oils, camphor, carvacrol, R(+) limonene, and linalool are major components in terms of content [106,107,108, 109,110,111]. The antibacterial action of various sage preparations was assessed for low-pressure mechanically separated meat in vacuum packaging stored at -18°C for nine months. Mechanically separated meat from chickens with the addition of sage extracts inhibited the growth of all groups of bacteria (mesophilic aerobic bacteria, psychrotrophic bacteria, Enterobacteriaceae, coliforms, and enterococci). The most effective antibacterial effect was exhibited by 0.1% sage essential oil treated groups [112,113,114,115,116,117]. Antibacterial effect of sage essential oil (0.625%) on survival of *Listeria monocytogenes* in Sous vides cook chill beef stored in refrigerated storage (two or 8 °C) for 28 days. A decrease of one log CFU / gram of *Listeria monocytogenes* was detected in sage essential oil treated groups compared to control at 2 °C. Exponential growth was observed from day 14, and decreased *Listeria monocytogenes* counts of one log CFU / gram were detected in sage essential oil-treated samples stored at 8°C [118,119,120,121,122,123].

Thyme

Thyme (*Thymus vulgaris*) is a representative herb used together with meat. Application of thyme in meat processing can elevate antioxidant, antibacterial, shelf-life extension, and sensory properties. In meat sausage, thyme essential oil inhibited 2.69 log Colony Forming Unit/gram of coagulase-positive *Staphylococcus* and 4.41 log Colony Forming Unit/gram of aerobic mesophilic bacteria, respectively, at a concentration of 0.95% by mixing with 1% (w/w) powdered beet juice. Sensory properties, odor, flavor, and overall acceptability improved [124,125,126,127,128,129]. The 1% thyme oil led to a reduction in *S. enterica* by three log Colony Forming Unit/gram during the margination process with lemon juice and 0.5% *Yucca schidigera* extract in raw chicken breast. Most compositions of thyme oil revealed 51.1% and 24.1% thymol and O-cymene, respectively. The antibacterial action of thyme may be due to additive or synergistic action with its major and/or minor components. Thymol and its synergistic effect with other phenolic compounds, such as carvacrol, p-cymene, and γ -terpinene, can change the permeability of bacterial cell walls, leading to cell death [130,131,132,133,134,135]. Thyme essential oil encapsulated with casein and maltodextrin was evaluated for its antibacterial potential in vitro and in situ (hamburger-like meat products). Encapsulated thyme essential oil showed the same minimum inhibitory concentration (0.1 mg/mL) against *Escherichia coli*, *S. Typhimurium*, *Staphylococcus aureus*, and *Listeria monocytogenes* as that of unencapsulated thyme essential. In treated groups with 1% (v/v) of encapsulated thyme essential oil for meat, *Escherichia coli* counts were decreased from 23 most probable number (MPN)/ gram to 0 MPN/ gram, which was similar to conventional preservative (sodium nitrate) used as a control until 14 days of refrigerated storage (4°C) [136,137,138,139,140,141].

Oregano

Oregano (*Origanum vulgare*) is regularly used in foods of the Mediterranean Sea area. Oregano essential oil has recognized antibacterial and antioxidant properties for extension of shelf-life. The antibacterial action of oregano was due to two bioactive polyphenols, thymol, and carvacrol

[142,143,144,145,146,147]. Oregano essential oil and its effect on the shelf-life of black wildebeest *Biceps femoris* muscles was investigated at 2.6°C. Components of oregano oil were thymol, carvacrol, ρ -cymene, β -caryophyllene, γ -terpinene, α -humulene, and α -pinene; among them, carvacrol (42.94%) and thymol (17.40%) were highest. Total viable counts and lactic acid bacteria reached the spoilage limit (seven log Colony Forming Unit/gram) after three days. Growth rates for total viable counts and lactic acid bacteria in the treated group were 40% higher than those in untreated groups [148,149,150,151,152,153]. The combinatorial effect of oregano essential oil with caprylic acid was studied in vacuum-packed minced beef. Addition of 0.2% oregano essential oil with 0.5% caprylic acid and 0.1% citric acid in minced beef reduced counts of lactic acid bacteria by 1.5 log Colony Forming Unit/gram in vacuum packaging. Cell counts of psychrotrophic bacteria and *Listeria monocytogenes* were reduced by more than 2.5 log Colony Forming Unit/gram at 3 °C for 10 days. Oregano essential oil inhibits the growth of bacteria by releasing volatile components during the drying process. The addition of oregano essential oil composed of carvacrol (64.5%), ρ -cymene (5.2%), and thymol (2.9%) inhibited *S. Enteritidis* and *Escherichia coli* in the beef drying process. For drying, a filter paper was soaked with oregano essential oil and placed in front of a fan of the drier. Beef samples were dried at 55 °C for 6 hours. Both bacteria (*S. Enteritidis* and *Escherichia coli*) were not detected after treatment with three mL of oregano essential oil [154,155,156, 157,158,159].

Chestnut

Castanea crenata was classified into the *Castanea* family and is a woody plant native to East Asia, including Korea and Japan. *Castanea sativa* is important for *Castanea* families and food resources of European areas for long periods. Chestnut shells contain abundant phenols and hydrolyzable tannins. Chestnut inner shell extracts using ethanol exhibited antimicrobial action against *C. jejuni* in chicken meat at a concentration of two mg/mL. Polyphenol and flavonoid contents of chestnut inner shell ethanol extracts were 532.96 ± 3.75 mg gallic acid/100 g and 12.28 ± 0.03 mg quercetin/100 g, respectively [160,161,162,163,164,165]. The action of chestnut extracts (*Castanea sativa*) on leaf, bur, and hull of beef patties under refrigerated conditions ($2 \pm 1^\circ\text{C}$) for 18 days to extend shelf-life. Inside chestnut extracts from leaf, bur, and hull, only leaf extract at a concentration of 1000 mg/kg had weak antimicrobial activity. Lactic acid bacteria and *Pseudomonas* spp. were reduced by 0.37 log Colony Forming Unit/gram and 0.33 log Colony Forming Unit/gram at seven days, respectively [166,167,168,169,170,171].

Grapefruit Seed Extract

Grapefruit Seed Extract is a by-product of *Citrus paradise*. Grapefruit Seed Extract contains various phenolic compounds and flavonoids, such as catechin, citric acid, naringenin, procyanidin, and epicatechin gallate. Grapefruit Seed Extract has been described to have a wide-ranging spectrum of antimicrobial, antiparasitic, and antifungal activities. Polyphenols in Grapefruit Seed Extract are unstable but can be chemically modified to become more stable using quaternary ammonium compounds, such as benzethonium chloride, during industrial procedure of commercial Grapefruit Seed Extract preparations [172,173,174,175,176,177]. Bacteriostatic effect of commercial Grapefruit Seed Extract (Citricidal) on sous-vide chicken

products against *Clostridium perfringens*. Cell numbers of *Clostridium perfringens* were consistently 2.5 log Colony Forming Unit/gram regardless of treatment or control groups until 9.5 h of stored at 19 °C; storage of control and 50 or 100 ppm Grapefruit Seed Extract treated groups at 25 °C for more than six hours resulted in fast growth rates of *Clostridium perfringens*, showing 2–3 log Colony Forming Unit/gram. Grapefruit Seed Extract concentrations at 200 ppm inhibited growth of *Clostridium perfringens* stored at 19 and 25°C. Active packaging system for inhibition of food-borne pathogens used mixed natural preservatives consisting of Grapefruit Seed Extract (80 mg/m²) with cinnamaldehyde (200 mg/m²) and nisin (60 mg/m²) was assessed for beef storage. Active packaging showed decreased contamination of psychrotrophic and anaerobic bacteria compared to control groups at 1–2 log Colony Forming Unit/gram. Packaged beef samples with mixed natural preservatives showed a decrease in *Listeria monocytogenes*, *Staphylococcus aureus*, and *C. jejuni* for 4.7 log Colony Forming Unit/gram, 0.81 log Colony Forming Unit/gram, and 3.1 log Colony Forming Unit/gram compared to wrapped packaging at 28 days of refrigerated storage, respectively. *C. jejuni* was observed below the detection limit after 21 days of storage [184,185,186,187,188,189].

Cinnamon

Cinnamon is a native plant in Asia that is acquired from the inner bark of the genus *Cinnamomum*. Cinnamon contains several active compounds, such as cinnamaldehyde, eugenol, cinnamyl acetate, L-borneol, β -caryophyllene, caryophyllene oxide, camphor, L-bornyl acetate, α -terpineol, α -cubebene, α -thujene, and terpinolene. Cinnamon (*Cinnamomum cassia*) essential oils could inhibit *L. monocytogenes* in ground beef at refrigerated (0 and 8 °C) and frozen (–18 °C) conditions. The concentration of five percent cinnamon essential oil decreased by 3.5–4.0 log Colony Forming Unit/gram of *Listeria monocytogenes* at 0 and 8 °C for seven days. Under frozen conditions, *Listeria monocytogenes* was reduced by 3.5–4.0 log Colony Forming Unit/gram over 60 days. Antibacterial effect and shelf-life extending activity were evaluated using a chitosan edible coating containing 0.6% cinnamon essential oil on roast duck slices under modified atmosphere packaging (30% carbon dioxide (CO₂)/70% nitrogen (N₂)) at storage at $2 \pm 2^\circ\text{C}$ for 21 days. Edible coating with cinnamon essential oil showed total viable counts reduced by one log Colony Forming Unit/gram compared to control after 14 days of storage. It is similar to the results of Enterobacteriaceae counts. The number of lactic acid bacteria decreased than that of control until day 7 of storage, but there was no significant difference from day 11 of storage. Growth of *Vibrio* spp. was delayed using edible coating with cinnamon essential oil within an earlier period of storage as a result of microbial diversity sequencing [196,197,198, 199, 200, 201].

Turmeric

Turmeric (*Curcuma longa* L.) has long been used as a flavor and color agent in food and traditional medicine to treat various diseases, mainly in South and East Asia. The main active compounds of turmeric originate from its constituents, called curcuminoids. Curcuminoids (curcumin, demethoxycurcumin, and bisdemethoxycurcumin) content of turmeric varies between about 2–9% based on its growth environments, such as cultivar, soil, and climatic conditions. The antibacterial effect of turmeric on chicken breast

meat was assessed for *Escherichia coli* and *Staphylococcus aureus* stored at 4 °C for two days. When 1% turmeric powder was added, no difference in *Staphylococcus aureus* counts was observed between turmeric-treated and control groups. In the case of *Escherichia coli*, a reduction of 0.2 log Colony Forming Unit/gram was observed, but this was not statistically significant [202,203,204,205,206,207]. Chicken meat was treated with turmeric powder and gamma irradiation to improve meat quality and stability. Total aerobic bacteria and coliforms were completely decontaminated with 3% turmeric powder and 2 kGy of gamma irradiation at 4 °C for 14 days. The microbial characteristics of edible coatings using turmeric starch and bovine gelatin were examined in frankfurter sausages. Edible coating was developed with a 5% (w/w) aqueous solution of turmeric starch and gelatin. Microbial growth of coated sausages stored at 5 °C for 20 days decreased by 2.21, 1.01, and 1.65 log Colony Forming Unit/gram for mesophilic bacteria, lactic acid bacteria, and psychotropic bacteria, respectively. At 10 °C, decreases were 1.57, 2.14, and 1.99 log Colony Forming Unit/gram for mesophilic bacteria, lactic acid bacteria, and psychotropic bacteria, respectively [208,209,210,211,212,213].

Plant-origin Antimicrobial Peptides

Plant origin Antimicrobial Peptides have been studied for their potential to inhibit different pathogenic bacteria, including food spoilage bacteria, food poisoning bacteria, mold, and yeast species. Antibacterial peptide Leg1 from chickpea legumin was reported in meat application of plant-origin Antimicrobial Peptides. Raw pork was pretreated with Leg1 and inoculated with *Escherichia coli* and *B. subtilis*. Bactericidal activity was measured at 37 °C for 16 hours. Minimum bactericidal concentrations of Leg1 on pork were 125 µM and 15.6 µM for *Escherichia coli* and *B. subtilis*, respectively. This was the same concentration as MBC of nisin, a bacteriocin from *Lactococcus lactis*, for tested strains. Antimicrobial Peptides from peas (11SGP) and red kidney beans (RBAH) were used to extend the shelf-life of raw buffalo meat. In laboratory media, Gram-positive (*L. monocytogenes*, *B. cereus*, and *Streptococcus pyogenes*) and Gram-negative (*Escherichia coli*, *Pseudomonas aeruginosa*, *Acinetobacter baumannii*) bacteria were inhibited by 11GSP (60 µg/mL) and Gram-negative bacteria by 60% and Gram-positive bacteria by 90%. RBAH (60 µg/mL) alleviated the growth of Gram-negative bacteria by 56% and Gram-positive bacteria by 85%. In buffalo meat, counts of mesophilic bacteria of 11SGP (400 µg/gram) and RBAG (400 µg/gram) treated groups decreased by 1.60 log Colony Forming Unit/gram and 1.94 log Colony Forming Unit/gram compared to control groups. Psychrophilic bacteria, 11SGP and RBAG reduced by 1.10 log Colony Forming Unit/gram and 1.47 log Colony Forming Unit/gram, respectively, after 15 d of refrigerated storage (4 °C) [172,173,174, 175,176].

Natural Preservatives from Animals and Their Application for Meat

Various antibacterial systems of animal sources are associated with defense mechanisms against external intruders. Preservatives derived from animal sources include lysozymes, lactoferrin, ovotransferrin, lactoperoxidase, Antimicrobial Peptides from livestock animals, and polysaccharides. Lysozyme can suppress several Gram-positive bacteria because of Lysozyme's distinctive ability to injure bacterial membranes by hydrolyzing 1,4-β-linkage between N acetyl D glucosamine and N acetyl muramic acid of

peptidoglycan in the bacterial membrane. Peptide-based antibacterial substances containing Antimicrobial Peptides from animal sources, ovotransferrin, and lactoferrin could influence cell membranes or synthesize Antimicrobial Peptides, peptides, and enzymes. The antibacterial mechanism of Antimicrobial Peptides is due to the attachment to the bacterial cell membrane and disturbs its integrity, resulting in cell lysis. Antimicrobial Peptides may also exert more complex activities that inhibit metabolic and translational systems. Ovotransferrin isolated from eggs increased the cell membrane permeability of Gram-positive and Gram-negative bacteria. Ovotransferrin destroyed cell membrane integrity, increased permeability of pathogen membranes, and induced morphological changes. Lactoferrin has antibacterial action related to large cationic patches present on the surface and iron impoverishment. Lactoferrin has an antibacterial effect only when in its iron-free state and iron-saturated lactoferrin has limited antimicrobial activity. Lactoperoxidase oxidizes sulfhydryl groups of proteins present in bacterial membranes, which could be injured by efflux of potassium ions, amino acids, peptides, and enzymes [177,178,179,180,181,182].

Lysozyme

Lysozyme (muramidase or N acetyl muramic hydrolase) is mainly extracted from hen egg whites and is known as an antimicrobial enzyme. Lysozyme is a glycoside hydrolase that hydrolyses linkages in peptidoglycan at Gram-positive bacterial cell walls. Lysozyme is composed of 129 amino acids, which contain disulfide bonds and tryptophan, tyrosine, and phenylalanine residues. Lysozyme, named Inovapure, has been used commercially against spoilage and foodborne pathogenic bacteria to prolong the shelf-life of raw and processed meat. Modified lysozyme, high hydrophobicity, and low hydrolytic activity compared to lysozyme monomer, at concentrations of 5%, exhibited low-microbial growth rates (total viable count 4.59 log CFU /cm²; molds and yeasts 2.17 log CFU /cm²) in pork surface with modified atmosphere packaging with composites of 50% O₂, 40% CO₂, and 10% N₂. Mixed antimicrobials consisting of lysozyme (250 ppm), nisin (250 ppm), and disodium ethylenediaminetetraacetic acid (EDTA) (20 mM) had antibacterial action against *Listeria monocytogenes*, total viable counts, Enterobacteriaceae, *Pseudomonas* spp., and lactic acid bacteria in ostrich meat patties with air and vacuum packaging. Mixed lysozyme preparations reduced *Listeria monocytogenes* below the official detection limit of the European Union (<2 log CFU /gram) in ostrich meat patties. Treated samples showed a decrease in total viable counts by one log CFU /gram after two days of storage and tended to increase thereafter. Enterobacteriaceae and *Pseudomonas* spp. were not affected by mixed antimicrobials in either packaging atmosphere, and a reduction in lactic acid bacteria was detected at two log CFU /gram. A combination of lysozyme with chitooligosaccharide presented a more effective antibacterial effect against Gram-negative bacteria than lysozyme alone. In minced mutton, a mixture of lysozyme and chitooligosaccharide led to the complete removal of 3–4 log CFU /gram of inoculated *Escherichia coli*, *Pseudomonas fluorescens*, and *B. cereus* during four hours at ambient temperature. *Staphylococcus aureus* was not completely eliminated but was reduced up to two log CFU/gram [183,184,185,186,187]

Ovotransferrin

Egg white contains 13% ovotransferrin (conalbumin), which is a monomeric 77.9 kDa glycoprotein comprised of 686 amino acid residues. Ovotransferrin contains N and C globular parts, each of which can reversibly bind Fe^{3+} and CO_3^{2-} . Ovotransferrin is the main constituent of the egg's defense system for bacteria, as it renders iron unusable for microbial growth within albumen. Antimicrobial action of ovotransferrin against *Escherichia coli* in fresh chicken breast involved in κ carrageenan film. Growth of *Escherichia coli* in fresh chicken breast wrapped with active film was 2.7 log CFU / gram by addition of 25 mg of ovotransferrin in combination with 5 mM EDTA. Ham models, 25 mg/mL of ovotransferrin with 100 mM sodium bicarbonate (NaHCO_3) did not show any antibacterial action against *Escherichia coli* O157:H7 and *Listeria monocytogenes* in commercial hams, whereas 25 mg/mL ovotransferrin with half percentage citric acid had bacteriostatic action against *Listeria monocytogenes* [188,189,190,191,192,193].

Lactoferrin

Lactoferrin, a glycoprotein that belongs to the transferrin protein family in milk and milk products as well as neutrophil granules and exocrine secretions in mammals, was able to bind iron within cells. The ability of this 80 kDa protein to control free iron levels contributes to its bacteriostatic and population health beneficial characteristics, such as stimulating bone growth, protecting intestinal epithelium, and promoting the immune system in animals. In ground beef, the application of active lactoferrin, immobilized lactoferrin with glycosaminoglycans, and solubilized in citrate/bicarbonate buffer systems at concentrations of three percent and five percentage resulted in two log CFU / gram reductions of *Escherichia coli* O157:H7 at 10 °C for nine days. The reduction of *S. Enteritidis* growth was 0.8 log CFU / gram when active lactoferrin concentration was increased to two-point-five percent. A single application of half percentage active lactoferrin reduced *Listeria monocytogenes* in beef, resulting in two log CFU / gram. Bovine lactoferrin (half mg) was tested against *Escherichia coli* O157:H7 and *P. fluorescens* inoculated on chicken with HPP treatments between 200 and 500 MPa for 10 min at 10 °C. As a result, *P. fluorescens* was decreased when lactoferrin was combined with HPP treatment at 300 MPa for 2.3 log CFU / gram additional reduction compared to only 300 MPa treatment on day 9. Additional reductions in *Escherichia coli* O157:H7 counts obtained by combined treatments remained below 0.5 log CFU/gram. [194,195,196,197,198]

Lactoperoxidase

Lactoperoxidase is a member of the peroxidase family. It is a ubiquitous active enzyme in bovine milk, which has antimicrobial action. Bovine lactoperoxidase is a glycoprotein that contains a peptide chain of 78.4 kDa and catalyzes the oxidation of thiocyanate ions (SCN^-) in lactoperoxidase, producing oxidizing products, such as hypothiocyanite and hypothiocyanous acid. Lactoperoxidase coated with alginate at concentrations of 2, 4, and 6% on the shelf-life of chicken breast filets. Chicken samples with an active coating of alginate and 6% lactoperoxidase showed a reduction of Enterobacteriaceae, *P. aeruginosa*, and aerobic mesophilic bacteria by five log CFU / gram, 4 log CFU / gram, and 2.5 log CFU / gram at 16 days of refrigerated storage, respectively. The antimicrobial action of lactoperoxidase was also assessed against *Listeria monocytogenes* and *S.*

Enteritidis in sliced dry-cured ham for 60 d at 8 °C treated with HPP at 450 MPa. The synergistic effect of lactoperoxidase and pressure was confirmed as *S. Enteritidis* decreased below the detection limit (one log CFU / gram). *Listeria monocytogenes*, synergistic effect reduced cell viability by 0.86 log CFU / gram compared with untreated samples at the end of storage. In beef, the effect of lactoperoxidase on the growth of inoculated pathogenic bacteria (four log CFU / gram) composed of *Staphylococcus aureus*, *Listeria monocytogenes*, *Escherichia coli* O157:H7, *S. Typhimurium*, *P. aeruginosa*, *Yersinia enterocolitica*, and indigenous microbiota was investigated. Pathogenic bacteria used in the experiment were reduced compared to control at a chilling regime (-1 to 12 °C) for 42 days. total aerobe and *Pseudomonas* spp. increased less in the lactoperoxidase-treated group than in the control group, but the antibacterial effect was not exhibited for anaerobes and lactic acid bacteria [199,200,201,202,203].

Livestock Animal Origin

Livestock animal-origin products have been used as a source of Livestock animal origin products. Among byproducts of livestock, blood, bones, collagen, gelatin, liver, lungs, placenta, skin, and visceral mass are important sources of Livestock animal origin products, as well as muscle parts. bovine cruor, a slaughterhouse byproduct containing mainly hemoglobin, broadly described as a rich source of fibrin proteins, was investigated for extraction of Livestock animal origin products. fraction named α 137–141 (polypeptide with five components, Thr Ser Lys Tyr Arg), a small (0.65 kDa), and hydrophilic Livestock animal origin products deviated from hemoglobin. The α 137–141 preservative (0.5%, w/w) had bacteriostatic action on total microbial population, coliform bacteria, yeasts, and molds at 4°C for 14d on beef. Livestock animal-origin products isolated from porcine leukocytes had antibacterial action on the proliferation of *Staphylococcus aureus* and *Escherichia coli* inoculated in ground meat (boneless ham) and sausage minces. The 20 μg / gram Livestock animal origin products decreased by 1.3 log CFU / gram of *Staphylococcus aureus* and 1.5 log CFU / gram of *Escherichia coli* in ground meat. It was also achieved that 160 μg / gram of Livestock animal origin products had the best inhibition and decreased in 3.9 log CFU / gram of *Staphylococcus aureus* and 3.3 log CFU / gram of *Escherichia coli* at 6 hours in ground meats. In sausage mince, Livestock animal origin products at concentrations of 160 μg / gram could decrease by three log CFU / g of *Staphylococcus aureus* and 2.7 log CFU / gram of *Escherichia coli* at 12 hours. After one day of storage, no visible colonies of *Staphylococcus aureus* or *Escherichia coli* were detected in sausage mince [203,204,205,206,207,208].

Natural Preservatives from Microorganisms and Their Application for Meat and its Products

Lactic acid bacteria strains secrete several bacterial growth inhibitory substances (organic acids, diacetyl, phenyl lactate, hydroxyphenyl lactate, cyclic dipeptides, hydroxy fatty acid, propionate, and hydrogen peroxide), bacteriocins (nisin, acidophilin, Bulgarian, Helvetica, lactacin, pediocin, plantarim, diplococcin, and bifidocin), and bacteriocin like inhibitory substances (Bacteriocin Like Inhibitory Substance), which exhibit antibacterial activity and can control spoilage and food-borne pathogens. various bacteriocins, commercial bacteriocin preparations have been applied

using nisin and pediocin. Bacteriocins are peptides or proteins with antibacterial and antifungal action that produce bacteria, mainly lactic acid bacteria. Compounds are considered potential natural preservatives because of their inhibitory action on food spoilage or food-borne pathogens. Lactic acid bacteria bacteriocins vary in accordance with molecular size, chemical structure, modifications during biosynthesis, presence of modified amino acid residues, and antimicrobial mechanisms. Lactic acid bacteria bacteriocins can be categorized into two major classes: class I (lanthionine containing antibiotics) with three subclasses (Ia, Ib, and Ic) and class II with four subclasses (IIa, IIb, IIc, and IId). Class I bacteriocins generally include 19–50 amino acid residues (<5 kDa) and are largely post-translationally modified, ensuring non-standard amino acids, such as lanthionine, β methylanthionine, and labyrinthine. Class I bacteriocins are further subdivided into class Ia (lantibiotics), class Ib (labyrinthopeptins), and class Ic (sanctibiotics). Class II bacteriocins comprise small, heat-stable, non-modified peptides (<10 kDa). It can be further subdivided into class IIa (pediocin-like bacteriocins), class IIb (non-modified bacteriocins with two or more peptides), class IIc (circular bacteriocins), and class IId (non-pediocin like bacteriocins). Pediocin-like bacteriocins (class IIa) can be regarded as the main subgroup among all classified Lactic acid bacteria bacteriocins. Class III bacteriocins are classified as high molecular weight (>30 kDa) and thermally unstable peptides. Class IV bacteriocins are large peptides complex with lipids or carbohydrates. Bacterial cell surface exhibits a negative charge because the anionic characteristics of the cell membrane consist of phosphatidylethanolamine, phosphatidylglycerol, lipopolysaccharide, lipoteichoic acid, and cardiolipin, and is generally captured by positively charged bacteriocins. Cationic charged groups of bacteriocins electrostatically interact with the anionic bacterial cell surface, while hydrophobic surfaces are attached to the membrane and traverse the lipid bilayer. Bacteriocins self-associate or polymerize to develop complexes after passing through the lipid bilayer. Bacteriocins induce cell death by increasing the permeability of bacterial membranes, forming pores that cause dissipation of proton motive force, exhaustion of ATP, and leakage of intracellular substrates. Gram-positive bacteria-origin bacteriocins only perform for Gram-positive bacteria and are not effective against Gram-negative bacteria because of their different membrane compositions and selective membrane permeability. Disadvantages could be compensated by mixing processing with other preservatives and the application of further preservation methods [209,210,211,212,213].

Nisin

Nisin is a most representative class I bacteriocin. Nisin is produced by several strains of *Lactococcus lactis*, a species that is widely used for dairy production. Nisin was first approved as a food preservative in the United Kingdom in the 1950s and is now widely used worldwide and is permitted in over 50 countries. The structure of nisin consists of a polypeptide with 34 amino acids, a 3.5 kDa molecular mass, and contains methylanthionine and lanthionine groups. Nisin has antimicrobial activities against a wide range of Gram-positive bacteria, including *Staphylococcus* spp., *Bacillus* spp., *Listeria* spp., and *Enterococcus* spp. Nisaplin is a typical commercial nisin formulation. Nisin could provide long-lasting bacteriostatic action on

pathogenic bacteria in beef jerky at room temperature. Shelf-life extensive effect of nisin in *B. cereus* inoculated with beef jerky. Beef jerky without nisin, counts of mesophilic bacteria, and *B. cereus* increasing is unlikely for beef jerky treated with nisin at 25°C for 60 days. *B. cereus* grew after three days in 100 IU nisin/gram treated groups and after 21 days in 500 IU/gram nisin-treated groups. Nisin-containing fermentate from *L. lactis* 537 strain was evaluated for inhibition of *Listeria monocytogenes* in ready-to-eat sliced ham. Addition of fermentate to ready-to-eat sliced ham led to an immediate decrease in *Listeria monocytogenes* counts from three log CFU/gram to below the detection limit stored at 4°C (20 CFU/gram). Nisin with cinnamaldehyde and grapefruit seed extract presented synergistic antibacterial action. It reduced counts of *Listeria monocytogenes* by three log CFU/gram in raw pork loin at 4°C for 12 hours. Minimum inhibitory concentration of nisin against *Listeria monocytogenes* was 250 ppm in laboratory media, but it was possible to reduce the concentration of 5–6 ppm against the growth of *Listeria monocytogenes* by mixing with natural antibacterial substances in pork [214,215,216,217,218,219].

Pediocin

Pediococcus spp., *Pediococcus acidilactici*, and *Pediococcus pentosaceus* are the main pediocin-producing strains. Pediocin was classified into bacteriocin group class IIa, characterized as small non-modified peptides (<5 kDa) comprising less than 50 amino acids. Remarkably, pediocin showed antimicrobial activity even at nanomolar concentrations. Using food-grade medicine-containing formulations are commercially available and marketed as ALTA 2341 and MicroGARD. Pediocin has been studied for inhibition of *Listeria* spp. for meat preservation methods. Antibacterial activities of pediocin PA 1 in frankfurters and *P. acidilactici* MCH14, pediocin PA 1 producing strain, in Spanish dry fermented sausages were assessed against *Listeria monocytogenes* and *Clostridium perfringens*. In frankfurters treated with 5000 bacteriocin units/mL of pediocin PA 1 produced by *P. acidilactici* MCH14, *Listeria monocytogenes* was reduced by 2 and 0.6 log CFU/gram after storage at 4°C for 60 days and at 15°C for one month, respectively. *Clostridium perfringens* decreased with 5000 BU/mL of pediocin PA 1 by two and 0.8 log CFU/gram after storage at 10°C for 60 d and at 15°C for one month, respectively. Growth of *Listeria monocytogenes* was inhibited by the pediocin-producing strain, *P. acidilactici* MCH14, in Spanish dry fermented sausages at two log CFU/gram compared to control. *bachA* 6111-2, pediocin from *P. acidilactici* HA 6111-2, was applied to Portuguese fermented meat sausage (Alheira) with HPP treatment (300 MPa, five min, 10°C) to inhibit *Listeria innocua*. Bacteriostatic effect was verified for high inoculation counts of *L. innocua* at 4°C for 60 days. Decreasing inoculated *L. innocua*, an antibacterial effect was observed below two log CFU/gram from day three of storage until the end of storage. Antibacterial activities of a mixed preparation containing pediocin from *Pediococcus pentosaceus* and *Murraya koenigii* (curry tree) berries in a raw goat meat emulsion at 4°C for 9 days. *L. innocua* was reduced for 4.1 log CFU/gram in treated samples concentrations at 8.3 mL pediocin/1000 grams of meat emulsion with 10% (v/w) *Murraya koenigii* berries extract at the end of storage. Total viable count and psychrophilic count were also observed to decrease in treated samples, 2.2

log CFU /gram and 1.6 log CFU / gram, respectively [220,221,222,223,224,225].

Sakacin

Sakacins, a class II bacteriocin, is mainly produced by *Lactobacillus sakei* or *Lactobacillus curvatus* strains. Commercial sakacin products are currently not presented. Compared to nisin and pediocin, sakacins have a relatively narrow antimicrobial spectrum, especially with effective inhibition against *Listeria* species. antibacterial effect of sakacin-producing strain, *L. sakei* CWBI B1365, and *L. curvatus* CWBI B28, on the fate of *Listeria monocytogenes* in raw beef and poultry. In refrigerated (5 °C) raw beef, *L. sakei* induced a decrease in *Listeria monocytogenes* concentration by 1.5 log CFU / gram after seven days to two log CFU / gram after 14 days, and below the detection limit at 21 days. addition of *L. curvatus* reduced *Listeria monocytogenes* to below the detection limit after seven days. In poultry, bacteriocin-producing strain did not affect inhibition of *Listeria monocytogenes*. It was assumed that type of meat may have influenced bacteriocin production by Lactic acid bacteria. Antibacterial activity of different bacteriocin preparations using sakacin Q produced by *L. curvatus* ACU 1 on meat surface was evaluated against *L. innocua*. freeze-dried reconstituted cell-free supernatant (3200 AU/mL) was effective for inhibition of *L. innocua* on meat surface, decreasing its bacterial cell number to the detection limit (<2 log CFU / gram) after two weeks of storage at 4–5 °C. adsorption of sakacin Q to meat products, main ingredients, meat proteins, and fat tissues did not affect its antibacterial activity [226,227,228,229,230,231].

Bacteriocin Inhibitory Substance

Bacteriocin Inhibitory substances are among the antimicrobial substances produced by bacteria and are not completely categorized in terms of amino acid composition, molecular size, and nucleotide sequence. Inside ready-to-eat pork ham, the antibacterial action of Bacteriocin Like Inhibitory Substance produced by *Pediococcus pentosaceus* American Type Culture Collection 43200 was assessed and compared with those of commercially available nisin preparations (Nisaplin). Bacteriocin-like inhibitory Substance showed effective antibacterial activity against *Listeria seeligeri* by 0.74 log CFU / gram in ready-to-eat ham stored at 4 °C after two days. A slight increase in *Listeria seeligeri* counts was detected in Bacteriocin-like inhibitory Substance-treated samples from six days to the end of storage. Nisaplin did not present any antibacterial effect for up to two days. After two days, Nisaplin started to induce a decrease in *Listeria seeligeri* counts throughout refrigerated storage. This might have been due to the higher sensitivity of Bacteriocin-like inhibitory Substance to residual proteases compared to nisin, so weakening its antibacterial effect. Bacteriocin Like Inhibitory Substance producing Lactic acid bacteria strains, *P. acidilactici* KTU05 7, *Pediococcus pentosaceus* KTU05 9, and *Listeria's sake* KTU05 6, were used to ferment plants (Jerusalem artichoke, *Helianthus tuberosus* L.), and 5% of fermented products were tested to inhibit foodborne pathogen at 18 °C for half day in ready to cook minced pork. *P. acidilactici* fermented product presented the highest antimicrobial activity compared to other strains. counts of *Escherichia coli*, *Enterococcus faecalis*, *Staphylococcus aureus*, and *Streptococcus* spp. were reduced by 5.53, 4.37, 4.86, and 3.84 log CFU /

gram, respectively, compared to control groups, suggesting that fermented product of Bacteriocin Like Inhibitory Substance producing strains showed an enhanced antibacterial effect. Bacteriocin- Inhibitory Substance obtained from *Enterococcus faecium* DB1 inhibited growth and formation of biofilms of *Clostridium perfringens* in chicken meat. The 2.5 mg/mL of DB1 Bacteriocin Like Inhibitory Substance suppressed the growth of *Clostridium perfringens* by 30%. *Clostridium perfringens* growth was inhibited by 50% at 5 mg/mL DB1 Bacteriocin Like Inhibitory Substance. Biofilm formation by *Clostridium perfringens* treated with 5 mg/mL DB1 Bacteriocin Like Inhibitory Substance was radically reduced by 90% at 4 °C for three days compared to control groups. The 2.5 mg/mL of DB1 Bacteriocin-like Inhibitory Substance also inhibited biofilm formation by *Clostridium perfringens* under the same conditions. Bacteriocin Like Inhibitory Substance could inhibit the formation of *Clostridium perfringens* biofilms on chicken surfaces due to its antibacterial effect [232,233,234,235,236,237].

Other Microorganism Sources

mytichitin CB peptide, which was isolated from blood lymphocytes of *Mytilus coruscus*, showed antibacterial action against Gram-positive bacteria and fungi. myricetin CB peptide expressed by *Pichia pastorisi* and applied to pork preservation methods. total viable counts of the treated group with 6 mg/L of mytichitin CB derived from *P. pastorisi* was reduced by 33% (1–2 log CFU / gram) compared to the control group after storage at 4°C for 5 days. Mytichitin CB effectively inhibited total bacterial growth during storage compared to groups treated with 50 mg/L of nisin. Mytichitin CB at 6 and 12 mg/L suppressed *Staphylococcus* spp. and *Escherichia* spp., respectively, with a reduction of 1–2 log Colony Forming units/gram, respectively. *Listeria* spp. And *Pseudomonas* spp. were not detected during storage, unlike control and nisin-treated groups. Hispidalin is a unique antimicrobial peptide derived from seeds of *Benincasa hispida* and has been shown to exhibit antimicrobial action against various bacteria. hispidalin expressed by *P. pastorisi* was used as a preservative for pork. Pork treated with 100 µg/mL hispidalin showed bacteriostatic action during the entire refrigerated storage period. total viable count of pork with 100 µg/mL hispidalin was one log CFU /gram decrease than that of the control group at 4°C for seven days [236,237, 238,239,240].

Conclusions

Meat is an important nutrient source due to its abundant protein content, essential amino acids, vitamins, and minerals. Meat is susceptible to contamination by food-borne pathogens and various spoilage bacteria because of its high water activity and important nutrient content. The application of preservatives is an indispensable element in livestock food processing to prevent food poisoning, delay spoilage, and extend their shelf-life. Industrial preservatives, commonly made up of chemicals, are not demanded by food customers because of their negative population health concerns. Natural preservatives derived from plants (rosemary, sage, chestnut, Grapefruit Seed Extract, and turmeric), animals (lysozyme, lactoferrin, lactoperoxidase, ovotransferrin, and others), and bacteria (organic acids, bacteriocins, and Bacteriocin Like Inhibitory Substance) have been explored as alternatives to chemical preservatives. versatility of natural preservatives compared to chemical preservatives is limited due to

production cost, standardization, insufficient toxicity studies, and negative sensory action on food. To compensate for these disadvantages, various applications have been studied for their synergistic effect with other natural preservatives with reduced application concentrations compared to single use, application of physical treatment such as gamma irradiation, high-pressure processing, drying, encapsulation, and the possibility of packaging materials. various natural preservatives and application methods to inhibit the growth of food-borne pathogens and spoilage bacteria in animal feed. Natural preservatives are expected to be in high demand due to consumer and industrial requests. Therefore, it is necessary to explore various applications of existing natural preservatives, while continuously searching for novel ones.

Conflicts of Interest

The author declares no conflicts of interest

References

- Shaltout, F.A, Riad, E.M, and AbouElhassan, Asmaa, A. (2017). prevalence Of Mycobacterium Tuberculosis In Imported cattle Offals And Its lymph Nodes. *Veterinary Medical Journal -Giza (VMJG)*. 63(2): 115-122.
- Papagianni M. (2003). Ribosomally synthesized peptides with antimicrobial properties: Biosynthesis, structure, function, and applications. *Biotechnol. Adv.* 21(6): 465-499.
- Shaltout, F.A, Riad,E.M , and Asmaa Abou-Elhassan. (2017). Prevalence Of Mycobacterium Spp. In Cattle Meat And Offal's Slaughtered In And Out Abattoir. *Egyptian Veterinary medical Association*. 77(2): 407-420.
- Meng D.-M, Sun S.-N, Shi L.-Y, Cheng L, Fan Z.-C. (2021). Application of antimicrobial peptide mytichitin-CB in pork preservation during cold storage. *Food Control*. 125: 108041.
- Abd Elaziz, O, Fatin S. Hassanin, Fahim A. Shaltout and Othman A. Mohamed. (2021). Prevalence of Some Foodborne Parasitic Affection in Slaughtered Animals in Local Egyptian Abattoir *Journal of Nutrition Food Science and Technology*. 6(2): 37-42.
- Del Olmo A, Calzada J, Nuñez M. (2012). Effect of lactoferrin and its derivatives, high hydrostatic pressure, and their combinations, on *Escherichia coli* O157:H7 and *Pseudomonas fluorescens* in chicken filets. *Innov. Food Sci. Emerg. Technol.* 13: 51–56.
- Abd Elaziz, O, Fatin, S Hassanin, Fahim, A Shaltout, Othman, A Mohamed. (2021). Prevalence of some zoonotic parasitic affections in sheep carcasses in a local abattoir in Cairo, Egypt. *Advances in Nutrition & Food Science*. 6(2): 25-31.
- Nieto-Lozano J.C, Reguera-Useros J.I, Peláez-Martínez M.d.C, Sacristán-Pérez-Minayo G, Gutiérrez-Fernández Á.J, de la Torre A.H. (2010). The effect of the pediocin PA-1 produced by *Pediococcus acidilactici* against *Listeria monocytogenes* and *Clostridium perfringens* in Spanish dry-fermented sausages and frankfurters. *Food Control*. 21: 679-685.
- Al Shorman,A.A. Shaltout, F.A. and hilat,N. (1999). Detection of certain hormone residues in meat marketed in Jordan.*Jordan University of Science and Technology, 1st International Conference on Sheep and goat Diseases and Productivity*. 23-25.
- Müller-Auffermann K, Grijalva F, Jacob F, Hutzler M. (2015). Nisin and its usage in breweries: A review and discussion. *J. Inst. Brew.* 121(3): 309-319.
- Ebeed Saleh, Fahim Shaltout, Essam Abd Elaal. (2021). Effect of some organic acids on microbial quality of dressed cattle carcasses in Damietta abattoirs, Egypt. *Damanhour Journal of Veterinary Sciences*. 5(2): 17-20.
- Cegielska-Radziejewska R, Szablewski T, Radziejewska-Kubzdela E, Tomczyk Ł, Biadała A, Leśnierowski G. (2021). The effect of modified lysozyme treatment on the microflora, physicochemical and sensory characteristics of pork packaged in preservative gas atmospheres. *Coatings*. 11(5): 488.
- Edris A, Hassanin, F. S. Shaltout, F.A, Azza H Elbaba and Nairoz M Adel. (2017). Microbiological Evaluation of Some Heat Treated Fish Products in Egyptian Markets.*EC Nutrition*. 12(3): 134-142.
- Elliot R.M, McLay J.C, Kennedy M.J, Simmonds R.S. (2004). Inhibition of foodborne bacteria by the lactoperoxidase system in a beef cube system. *Int. J. Food Microbiol.* 91(1): 73-81.
- Edris,A, Hassan,M.A, Shaltout,F.A. and Elhosseiny, S. (2013). Chemical evaluation of cattle and camel meat.*BENHA VETERINARY MEDICAL JOURNAL*. 25(2): 145-150.
- Rao M.S, Chander R, Sharma A. (2008). Synergistic effect of chitooligosaccharides and lysozyme for meat preservation. *LWT*. 41(10):1995-2001.
- Edris,A.M, Hassan,M.A, Shaltout,F.A. and Elhosseiny , S. (2013). Detection of E.coli and Salmonella organisms in cattle and camel meat. *BENHA VETERINARY MEDICAL JOURNAL*. 25(2): 198-204.
- Przybylski R, Firdaous L, Chataigne G, Dhulster P, Nedjar N. (2016). Production of an antimicrobial peptide derived from slaughterhouse by-product and its potential application on meat as preservative. *Food Chem*. 211:306-313.
- Edris A.M. Hemmat, M. I, Shaltout F.A. Elshater M.A. Eman F.M.I. (2012). STUDY ON INCIPIENT SPOILAGE OF CHILLED CHICKEN CUTS-UP. *BENHA VETERINARY MEDICAL JOURNAL*. 23(1): 81-86.
- Lee J.S, Park S.W, Lee H.B, Kang S.S. (2021). Bacteriocin-like inhibitory substance (BLIS) activity of *Enterococcus faecium* DB1 against biofilm formation by *Clostridium perfringens*. *Probiotics Antimicrob. Proteins*. 13:1452–1457.
- Edris A.M. Hemmat, M.I. Shaltout, F.A. Elshater M.A, Eman, F.M.I. (2012). CHEMICAL ANALYSIS OF CHICKEN MEAT WITH RELATION TO ITS QUALITY. *BENHA VETERINARY MEDICAL JOURNAL*. 23(1): 88-93.
- Rolfe V, Mackonochie M, Mills S, McLennan E. (2020). Turmeric/curcumin and health outcomes: A meta-review of systematic reviews. *Eur. J. Integr. Med.* 40: 101252.

23. Edris, A.M. Shaltout, F.A. and Abd Allah, A.M. (2005). Incidence of *Bacillus cereus* in some meat products and the effect of cooking on its survival. *Zag. Vet. J.* 33 (2):118-124.
24. Dong A, Malo A, Leong M, Ho V.T.T, Turner M.S. (2021). Control of *Listeria monocytogenes* on ready-to-eat ham and fresh cut iceberg lettuce using a nisin containing *Lactococcus lactis fermentate*. *Food Control.* 119: 107420.
25. Edris, A.M. Shaltout, F.A. and Arab, W.S. (2005). Bacterial Evaluation of Quail Meat. *Benha Vet. Med.J.* 16(1): 1-14.
26. Shwaiki L.N, Lynch K.M, Arendt E.K. (2021). Future of antimicrobial peptides derived from plants in food application—A focus on synthetic peptides. *Trends Food Sci. Technol.* 112: 312-324.
27. Edris, A.M. Shaltout, F.A. Salem, G.H. and El-Toukhy,E.I. (2011). Incidence and isolation of *Salmonellae* from some meat products. *Benha University ,Faculty of Veterinary Medicine , Fourth Scientific Conference 25-27th May 2011Veterinary Medicine and Food Safety)benha , Egypt.* 172-179.
28. Jaspal M.H, Ijaz M, Haq H.A.u, Yar M.K, Asghar B, Manzoor A, Badar I.H, Ullah S, Islam M.S, Hussain J. (2021). Effect of oregano essential oil or lactic acid treatments combined with air and modified atmosphere packaging on the quality and storage properties of chicken breast meat. *LWT.* 146(5): 111459.
29. Edris AA, Hassanin, F. S Shaltout, F.A, Azza H Elbaba and Nairoz M Adel. (2017). Microbiological Evaluation of Some Heat Treated Fish Products in Egyptian Markets. *EC Nutrition.* 12(3): 134-142.
30. Wang F.-S. (2003). Effect of antimicrobial proteins from porcine leukocytes on *Staphylococcus aureus* and *Escherichia coli* in comminuted meats. *Meat Sci.* 65(1): 615-621.
31. Edris, A.M. Shaltout, F.A. Salem, G.H. and El-Toukhy,E.I. (2011). Plasmid profile analysis of *Salmonellae* isolated from some meat products. *Benha University ,Faculty of Veterinary Medicine , Fourth Scientific Conference 25-27th May 2011Veterinary Medicine and Food Safety)194-201 benha , Egypt.*1:172-178.
32. Seol K.H, Lim D.G, Jang A, Jo C, Lee M. (2009). Antimicrobial effect of kappa-carrageenan-based edible film containing ovotransferrin in fresh chicken breast stored at 5 degrees C. *Meat Sci.* 83(3): 479-483.
33. Ragab A, Abobakr M. Edris, Fahim A.E. Shaltout, Amani M. Salem. (2022). Effect of titanium dioxide nanoparticles and thyme essential oil on the quality of the chicken fillet. *BENHA VETERINARY MEDICAL JOURNAL.* 41(2): 38-40.
34. Anastasio A, Marrone R, Chirollo C, Smaldone G, Attouchi M, Adamo P, Sadok S, Pepe T. (2014). Swordfish steaks vacuum-packed with *Rosmarinus officinalis*. *Ital. J. Food Sci.* 26(4): 390-397.
35. Woraprayote W, Malila Y, Sorapukdee S, Swetwivathana A, Benjakul S, Visessanguan W. (2016). Bacteriocins from lactic acid bacteria and their applications in meat and meat products. *Meat Sci.* 120: 118–132.
36. Hassan, M.A, Shaltout, F. A, Arfa M.M, Mansour A.H and Saudi, K. R. (2013). BIOCHEMICAL STUDIES ON RABBIT MEAT RELATED TO SOME DISEASES. *BENHA VETERINARY MEDICAL JOURNAL.* 25(1):88-93.
37. Mastromatteo M, Lucera A, Sinigaglia M, Corbo M.R. (2010). Synergic antimicrobial activity of lysozyme, nisin, and EDTA against *Listeria monocytogenes* in ostrich meat patties. *J. Food Sci.* 75(7): 422-429.
38. Hassan, M.A and Shaltout, F.A. (1997). Occurrence of Some Food Poisoning Microorganisms In Rabbit Carcasses *Alex.J.Vet.Science.* 13(1): 55-62.
39. Lee N.-K, Paik H.-D. (2021). Prophylactic effects of probiotics on respiratory viruses including COVID-19: A review. *Food Sci. Biotechnol.* 30(6): 773–781.
40. Hassan M, Shaltout FA* and Saqur N. (2020). Histamine in Some Fish Products. *Archives of Animal Husbandry & Dairy Science.* 2(1): 1-3.
41. Wu T, Wu C, Fu S, Wang L, Yuan C, Chen S, Hu Y. (2017). Integration of lysozyme into chitosan nanoparticles for improving antibacterial activity. *Carbohydr. Polym.* 155: 192-200.
42. Hassan, M.A and Shaltout, F.A. (2004). Comparative Study on Storage Stability of Beef, Chicken meat, and Fish at Chilling Temperature. *Alex.J.Vet.Science.* 20(21): 81-86.
43. Stimbirys A, Bartkiene E, Siugzdaite J, Augeniene D, Vidmantiene D, Juodeikiene G, Maruska A, Stankevicius M, Cizeikiene D. (2015). Safety and quality parameters of ready-to-cook minced pork meat products supplemented with *Helianthus tuberosus* L. tubers fermented by BLIS producing lactic acid bacteria. *J. Food Sci. Technol.* 52(7): 4306-4314.
44. Hassan, M.A Shaltout, F.A. Arafa, M.M. Mansour, A.H. and Saudi, K.R. (2013). Biochemical studies on rabbit meat related to some diseases. *Benha Vet. Med.J.* 25(1): 88-93.
45. Lee D, Heinz V, Knorr D. (2003). Effects of combination treatments of nisin and high-intensity ultrasound with high pressure on the microbial inactivation in liquid whole egg. *Innov. Food Sci. Emerg. Technol.* 4(4): 387–393.
46. Hassan, M.A Shaltout, F.A. Maarouf, A.A. and El-Shafey, W.S. (2014). Psychrotrophic bacteria in frozen fish with special reference to *Pseudomonas* species. *Benha Vet. Med.J.* 27(1): 78-83.
47. Lee N.K, Kim H.W, Lee J.Y, Ahn D.U, Kim C.J, Paik H.D. (2015). Antimicrobial effect of nisin against *Bacillus cereus* in beef jerky during storage. *Korean J. Food Sci. Anim. Resour.* 35(2): 272–276.
48. Hassan, M.A Shaltout, F.A. Arafa, M.M. Mansour, A.H. and Saudi, K.R. (2013). Bacteriological studies on rabbit meat related to some diseases *Benha Vet. Med.J.* 25(1): 88-93.
49. Yousefi M, Farshidi M, Ehsani A. (2018). Effects of lactoperoxidase system-alginate coating on chemical, microbial, and sensory properties of chicken breast fillets during cold storage. *J. Food Saf.* 38(3):12449.
50. Hassanin, F. S Hassan, M.A, Shaltout, F.A, Nahla A. Shawqy and 2Ghada A. Abd-Elhameed. (2017). Chemical criteria of chicken meat. *BENHA VETERINARY MEDICAL JOURNAL.* 33(2): 457-464.
51. Delves-Broughton J. (2012). Natural antimicrobials as additives and ingredients for the preservation of foods and beverages. In: Baines D,

- Seal R, editors. Natural Food Additives, Ingredients and Flavourings. 1st ed. Woodhead Publishing Series in Food Science, Technology and Nutrition; Cambridge, UK: 127-161.
52. Hassanin, F. S Hassan, M.A. Shaltout, F.A. and Elrais-Amina, M. (2014). CLOSTRIDIUM PERFRINGENS IN VACUUM PACKAGED MEAT PRODUCTS. BENHA VETERINARY MEDICAL JOURNAL. 26(1): 49-53.
53. Kumar Y, Kaur K, Shahi A.K, Kairam N, Tyagi S.K. (2017). Antilisterial, antimicrobial and antioxidant effects of pediocin and *Murraya koenigii* berry extract in refrigerated goat meat emulsion. LWT. 79(4):135-144.
54. Hassanien, F.S. Shaltout, F.A. Fahmey, M.Z. and Elsukkary, H.F. (2020). Bacteriological quality guides in local and imported beef and their relation to public health. Benha Veterinary Medical Journal. 39(1): 125-129.
55. Montville T.J, Bruno M.E.C. (1994). Evidence that dissipation of proton motive force is a common mechanism of action for bacteriocins and other antimicrobial proteins. Int. J. Food Microbiol. 24(1-2): 53-74.
56. Hassanin, F. S Shaltout, F.A. and , Mostafa E.M. (2013). Parasitic affections in edible offal. Benha Vet. Med.J. 25(1): 46-55.
57. Kotra V.S.R, Satyabanta L, Goswami T.K. (2019). A critical review of analytical methods for determination of curcuminoids in turmeric. J. Food Sci. Technol. 56(12): 5153–5166.
58. Hassanin, F. S Shaltout, F.A, Lamada, H.M, Abd Allah, E.M. (2011). THE EFFECT OF PRESERVATIVE (NISIN) ON THE SURVIVAL OF LISTERIA MONOCYTOGENES. BENHA VETERINARY MEDICAL JOURNAL. 1: 141-145.
59. Ma B, Guo Y, Fu X, Jin Y. (2020). Identification and antimicrobial mechanisms of a novel peptide derived from egg white ovotransferrin hydrolysates. LWT. 131(1-3): 109720.
60. Khattab, E, Fahim Shaltout and Islam Sabik. (2021). Hepatitis A virus related to foods. BENHA VETERINARY MEDICAL JOURNAL. 40(1): 174-179.
61. Sedighi R, Zhao Y, Yerke A, Sang S. (2015). Preventive and protective properties of rosemary (*Rosmarinus officinalis* L.) in obesity and diabetes mellitus of metabolic disorders: A brief review. Curr. Opin. Food Sci. 2:58–70.
62. Saad M. Saad, Fahim A. Shaltout, Amal A. A. Farag Hashim F. Mohammed. (2022). Organophosphorus Residues in Fish in Rural Areas. Journal of Progress in Engineering and Physical Science. 1(1): 27-31.
63. Dortu C, Huch M, Holzapfel W.H, Franz C.M, Thonart P. (2008). Anti-listerial activity of bacteriocin-producing *Lactobacillus curvatus* CWBI-B28 and *Lactobacillus sakei* CWBI-B1365 on raw beef and poultry meat. Lett. Appl. Microbiol. 47(6): 581-586.
64. Saif, M, Saad S.M, Hassanin, F. S Shaltout FA, Marionette Zaghoul. (2019). Molecular detection of enterotoxigenic *Staphylococcus aureus* in ready-to-eat beef products. Benha Veterinary Medical Journal. 37(1): 7-11.
65. Massantini R, Moscetti R, Frangipane M.T. (2021). Evaluating progress of chestnut quality: A review of recent developments. Trends Food Sci. Technol. 113: 245-254.
66. Saif, M, Saad S.M, Hassanin, F. S Shaltout, F.A, Marionette Zaghoul. (2019). Prevalence of methicillin-resistant *Staphylococcus aureus* in some ready-to-eat meat products. Benha Veterinary Medical Journal. 37(1): 12-15.
67. Papagianni M, Anastasiadou S. (2009). Pediocins: The bacteriocins of *Pediococci*. Sources, production, properties and applications. Microb. Cell Factories. 8:3.
68. Farag, A. A, Saad M. Saad¹, Fahim A. Shaltout¹, Hashim F. Mohammed. (2023). Studies on Pesticides Residues in Fish in Menofia Governorate. Benha Journal of Applied Sciences. 8(5): 323-330.
69. Castro S, Silva J, Casquete R, Queirós R, Saraiva J, Teixeira P. (2018). Combined effect of pediocin bacHA-6111-2 and high hydrostatic pressure to control *Listeria innocua* in fermented meat sausage. Int. Food Res. J. 25(2):553–560.
70. Farag, A. A, Saad M. Saad¹, Fahim A. Shaltout¹, Hashim F. Mohammed. (2023). Organochlorine Residues in Fish in Rural Areas. Benha Journal of Applied Sciences. 8(5): 331-336.
71. Kumariya R, Garsa A.K, Rajput Y.S, Sood S.K, Akhtar N, Patel S. (2019). Bacteriocins: Classification, synthesis, mechanism of action and resistance development in food spoilage causing bacteria. Microb. Pathog. 128:171-177.
72. Shaltout, F.A, Mona N. Hussein, Nada Kh. Elsayed. (2023). Histological Detection of Unauthorized Herbal and Animal Contents in Some Meat Products. Journal of Advanced Veterinary Research. 13(2): 157-160.
73. Marchese A, Orhan I.E, Daglia M, Barbieri R, Di Lorenzo A, Nabavi S.F, Gortzi O, Izadi M, Nabavi S.M. (2016). Antibacterial and antifungal activities of thymol: A brief review of the literature. Food Chem. 210: 402–414.
74. Shaltout, F. A, Heikal, G. I, Ghanem, A. M. (2022). Mycological quality of some chicken meat cuts in Gharbiya governorate with special reference to *Aspergillus flavus* virulent factors. benha veteriv medical journal veterinary. 42(1): 12-16.
75. De Azevedo P.O.d.S, Converti A, Gierus M, de Souza Oliveira R.P. (2019). Antimicrobial activity of bacteriocin-like inhibitory substance produced by *Pediococcus pentosaceus*: From shake flasks to bioreactor. Mol. Biol. Rep. 46(1):461-469.
76. Shaltout, F.A, Ramadan M. Salem, Eman M. Eldiasty, Fatma A. Diab. (2022). Seasonal Impact on the Prevalence of Yeast Contamination of Chicken Meat Products and Edible Giblets. Journal of Advanced Veterinary Research. 12(5): 641-644.
77. Shahnawaz M, Soto C. (2012). Microcin amyloid fibrils A are reservoir of toxic oligomeric species. J. Biol. Chem. 287(15):11665–11676.
78. Shaltout, F.A, Abdelazez Ahmed Helmy Barr and Mohamed Elsayed Abdelaziz. (2022). Pathogenic Microorganisms in Meat Products.

- Biomedical Journal of Scientific & Technical Research. 41(4): 32836-32843.
79. De Alba M, Bravo D, Medina M. (2015). Inactivation of *Listeria monocytogenes* and *Salmonella Enteritidis* in dry-cured ham by combined treatments of high pressure and the lactoperoxidase system or lactoferrin. *Innov. Food Sci. Emerg. Technol.* 31: 54-59.
80. Shaltout, F.A, Thabet, M.G. and Koura, H.A. (2017). Impact of Some Essential Oils on the Quality Aspect and Shelf Life of Meat. *J Nutr Food Sci.* 7(6): 100647.
81. Moon S.H, Paik H.D, White S, Daraba A, Mendonca A.F, Ahn D.U. (2011). Influence of nisin and selected meat additives on the antimicrobial effect of ovotransferrin against *Listeria monocytogenes*. *Poult. Sci.* 90(11): 2584–2591.
82. Shaltout, F.A, Islam Z. Mohammed², El -Sayed A. Afify. (2020). Bacteriological profile of some raw chicken meat cuts in Ismailia city, Egypt. *Benha Veterinary Medical Journal.* 39: 11-15.
83. Ko K.Y, Mendonca A.F, Ahn D.U. (2008). Influence of zinc, sodium bicarbonate, and citric acid on the antibacterial activity of ovotransferrin against *Escherichia coli* O157:H7 and *Listeria monocytogenes* in model systems and ham. *Poult. Sci.* 87(12): 2660–2670.
84. Shaltout, F.A, Islam, Z. Mohammed², El -Sayed A. Afify. (2020). Detection of *E. coli* O157 and *Salmonella* species in some raw chicken meat cuts in Ismailia province, Egypt. *Benha Veterinary Medical Journal.* 39: 101-104.
85. Meng D.-M, Sun X.-Q, Sun S.-N, Li W.-J, Lv Y.-J, Fan Z.-C. (2020). The potential of antimicrobial peptide Hispidalin application in pork preservation during cold storage. *J. Food Process. Preserv.* 44: 14443.
86. Shaltout, F.A, E.M. El-diasty and M. A. Asmaa- Hassan. (2020). HYGIENIC QUALITY OF READY TO EAT COOKED MEAT IN RESTAURANTS AT *Cairo*. *Journal of Global Biosciences.* 8(12): 6627-6641.
87. Giansanti F, Panella G, Leboffe L, Antonini G. (2016). Lactoferrin from milk: Nutraceutical and pharmacological properties. *Pharmaceuticals.* 9(4): 61.
88. Shaltout, F.A, Marrionet Z. Nasief, L. M. Lotfy, Bossi T. Gamil. (2019). Microbiological status of chicken cuts and its products. *Benha Veterinary Medical Journal.* 37: 57-63.
89. Vasconcelos N, Croda J, Simionatto S. (2018). Antibacterial mechanisms of cinnamon and its constituents: A review. *Microb. Pathog.* 120: 198–203.
90. Shaltout, F.A. (2019). Poultry Meat. *Scholarly Journal of Food and Nutrition.* 2(2): 1-2.
91. Mei J, Ma X, Xie J. (2019). Review on natural preservatives for extending fish shelf life. *Foods.* 8(10): 490.
92. Shaltout, F.A. (2019). Food Hygiene and Control. *Food Science and Nutrition Technology.* 4(5): 1-2.
93. Yuan S, Yin J, Jiang W, Liang B, Pehkonen S, Choong C. (2013). Enhancing antibacterial activity of surface-grafted chitosan with immobilized lysozyme on bioinspired stainless steel substrates. *Colloids Surf. B.* 106: 11–21.
94. Hassanin, F. S Shaltout, F.A, Seham N. Homouda and Safaa M. Arakeeb. (2019). Natural preservatives in raw chicken meat. *Benha Veterinary Medical Journal.* 37(1): 41-45.
95. Xu M.M, Kaur M, Pillidge C.J, Torley P.J. (2021). Microbial biopreservatives for controlling the spoilage of beef and lamb meat: Their application and effects on meat quality. *Crit. Rev. Food Sci. Nutr.* 62(3):1–35.
96. Hazaa,W, Shaltout, F.A, Mohamed El-Shate. (2019). Prevalence of some chemical hazards in some meat products. *Benha Veterinary Medical Journal.* 37(2): 32-36.
97. Rivas F.P, Castro M.P, Vallejo M, Marguet E, Campos C.A. (2014). Sakacin Q produced by *Lactobacillus curvatus* ACU-1: Functionality characterization and antilisterial activity on cooked meat surface. *Meat Sci.* 97(4):475-479.
98. Hazaa,W, Shaltout, F.A, Mohamed El-Shater. (2019). Identification of Some Biological Hazards in Some Meat Products. *Benha Veterinary Medical Journal* 37(2): 27-31.
99. De Azevedo P.O.S, Mendonca C.M.N, Seibert L, Dominguez J.M, Converti A, Gierus M, Oliveira R.P.S. (2020). Bacteriocin-like inhibitory substance of *Pediococcus pentosaceus* as a biopreservative for *Listeria* sp. control in ready-to-eat pork ham. *Braz. J. Microbiol.* 51(3): 949-956.
100. Gaafar,R, Hassanin, F. S Shaltout, F.A, Marionette Zaghloul. (2019). Molecular detection of enterotoxigenic *Staphylococcus aureus* in some ready to eat meat-based sandwiches. *Benha Veterinary Medical Journal.* 37(2): 22-26.
101. Parada J.L, Caron C.R, Medeiros A.B.P, Soccol C.R. (2007). Bacteriocins from lactic acid bacteria: Purification, properties and use as biopreservatives. *Braz. Arch. Biol. Technol.* 50(3): 512-542.
102. Gaafar,R, Hassanin, F. S Shaltout, F.A, Marionette Zaghloul.(2019). Hygienic profile of some ready to eat meat product sandwiches sold in Benha city, Qalubiyah Governorate, Egypt. *Benha Veterinary Medical Journal.* 37 (1) 16-21.
103. Galvez A, Abriouel H, Benomar N, Lucas R. (2010). Microbial antagonists to food-borne pathogens and biocontrol. *Curr. Opin. Biotechnol.* 21(2): 142–148.
104. Saad S.M, Shaltout, F.A, Nahla A Abou Elroos, Saber B El-nahas. (2019). Antibacterial Effect of Some Essential Oils on Some Pathogenic Bacteria in Minced Meat. *J Food Sci Nutr Res.* 2(1): 013-021.
105. Yu H.H, Song M.W, Song Y.J, Lee N.K, Paik H.D. (2019). Antibacterial effect of a mixed natural preservative against *Listeria monocytogenes* on lettuce and raw pork loin. *J. Food Prot.* 82(11): 2001–2006.
106. Tosati J.V, Messias V.C, Carvalho P.I.N, Rodrigues Pollonio M.A, Meireles M.A.A, Monteiro A.R. (2017). Antibacterial effect of edible coating blend based on turmeric starch residue and gelatin applied onto fresh frankfurter sausage. *Food Bioproc. Technol.* 10(1): 2165–2175.

107. Saad S.M, Hassanin, F. S. Shaltout, F.A, Marionette Z Nassif, Marwa Z Seif. (2019). Prevalence of Methicillin-Resistant Staphylococcus Aureus in Some Ready-to-Eat Meat Products. *American Journal of Biomedical Science & Research*. 4(6): 460-464.
108. Gogliettino M, Balestrieri M, Ambrosio R.L, Anastasio A, Smaldone G, Proroga Y.T, Moretta R, Rea I, De Stefano L, Agrillo B. (2020). Extending the shelf-life of meat and dairy products via PET-modified packaging activated with the antimicrobial peptide MTP1. *Front. Microbiol*. 10: 2963.
109. Shaltout, Fahim. (2019). Pollution of Chicken Meat and Its Products by Heavy Metals. *Research and Reviews on Healthcare: Open Access Journal*. 4(3): 381-382.
110. Chen X, Chen W, Lu X, Mao Y, Luo X, Liu G, Zhu L, Zhang Y. (2021). Effect of chitosan coating incorporated with oregano or cinnamon essential oil on the bacterial diversity and shelf life of roast duck in modified atmosphere packaging. *Food Res. Int*. 147: 110491.
111. Shaltout, F. A. E.M EL-diahy M. S. M Mohamed. (2018). Effects of chitosan on quality attributes fresh meat slices stored at 4 C. *BENHA VETERINARY MEDICAL JOURNAL*. 35(2): 157-168.
112. Lazzaro B.P, Zasloff M, Rolff J. (2020). Antimicrobial peptides: Application informed by evolution. *Science*. 368(6490): 5480.
113. Shaltout, F. A. and Abdel-Aziz. (2004). Salmonella enterica serovar Enteritidis in poultry meat and their epidemiology. *Vet. Med. J. Giza*. 52(3): 429-436.
114. Borrajo P, Pateiro M, Barba F.J, Mora L, Franco D, Toldrá F, Lorenzo J.M. (2019). Antioxidant and antimicrobial activity of peptides extracted from meat by-products: A review. *Food Anal. Methods*. 12: 2401–2415.
115. Shaltout, F.A, Hala F El-Shorah, Dina I El Zahaby, Lamiaa M Lotfy. (2018). Bacteriological Profile of Chicken Meat Products. *SciFed Food & Dairy Technology Journal*. 1(3): 83-90.
116. Cegielka A, Hac-Szymanczuk E, Piwowarek K, Dasiewicz K, Slowinski M, Wronska K. (2019). The use of bioactive properties of sage preparations to improve the storage stability of low-pressure mechanically separated meat from chickens. *Poult. Sci*. 98(10): 5045–5053.
117. Eslamloo K, Falahatkar B, Yokoyama S. (2012). Effects of dietary bovine lactoferrin on growth, physiological performance, iron metabolism and non-specific immune responses of Siberian sturgeon *Acipenser baeri*. *Fish Shellfish Immunol*. 32(6): 976–985.
118. Shaltout, F.A, Mohamed, A.H. El-Shater, Wafaa Mohamed Abd El-Aziz. (2015). Bacteriological assessment of Street Vended Meat Products sandwiches in kalyobia Governorate. *BENHA VETERINARY MEDICAL JOURNAL*. 28(2): 58-66.
119. El-Saadony M.T, Abd El-Hack M.E, Swelum A.A, Al-Sultan S.I, El-Ghareeb W.R, Hussein E.O.S, Ba-Awadh H.A, Akl B.A, Nader M.M. (2021). Enhancing quality and safety of raw buffalo meat using the bioactive peptides of pea and red kidney bean under refrigeration conditions. *Ital. J. Anim. Sci*. 20: 762–776.
120. Shaltout, F.A, Mohamed A El shatter and Heba M Fahim. (2019). Studies on Antibiotic Residues in Beef and Effect of Cooking and Freezing on Antibiotic Residues Beef Samples. *Scholarly Journal of Food and Nutritionm*. 2(1): 1-4.
121. Heymich M.L, Srirangan S, Pischetsrieder M. (2021). Stability and activity of the antimicrobial peptide Leg1 in solution and on meat and its optimized generation from chickpea storage protein. *Foods*. 10(6): 1192.
122. Shaltout FA, Zakaria IM and Nabil ME. (2018). Incidence of Some Anaerobic Bacteria Isolated from Chicken Meat Products with Special Reference to Clostridium perfringens. *Nutrition and Food Toxicology*. 2(5): 429-438.
123. Khaleque M.A, Keya C.A, Hasan K.N, Hoque M.M, Inatsu Y, Bari M.L. (2016). Use of cloves and cinnamon essential oil to inactivate *Listeria monocytogenes* in ground beef at freezing and refrigeration temperatures. *LWT*. 74(4): 219–223.
124. Shaltout FA, Ahmed A A Maarouf and Mahmoud ES Elkhoully. (2017). Bacteriological Evaluation of Frozen Sausage. *Nutrition and Food Toxicology*. 1(5): 174-185.
125. Stojanović-Radić Z, Pejčić M, Joković N, Jokanović M, Ivić M, Šojić B, Škaljac S, Stojanović P, Mihajilov-Krstev T. (2018). Inhibition of *Salmonella Enteritidis* growth and storage stability in chicken meat treated with basil and rosemary essential oils alone or in combination. *Food Control*. 90(4): 332–343.
126. Shaltout FA, El-Toukhy EI and Abd El-Hai MM. (2019). Molecular Diagnosis of Salmonellae in Frozen Meat and Some Meat Products. *Nutrition and Food Technology Open Access*. 5(1): 1-6.
127. Moura-Alves M, Gouveia A.R, de Almeida J.M.M.M, Monteiro-Silva F, Silva J.A, Saraiva C. (2020). Behavior of *Listeria monocytogenes* in beef Sous vide cooking with *Salvia officinalis* L. essential oil, during storage at different temperatures. *LWT*. 132: 109896.
128. Shaltout, F.A, A.M.Ali and S.M.Rashad. (2016). Bacterial Contamination of Fast Foods. *Benha Journal of Applied Sciences (BJAS)*. 1 (2): 45-51.
129. Yu H.H, Kim Y.J, Park Y.J, Shin D.-M, Choi Y.-S, Lee N.-K, Paik H.-D. (2020). Application of mixed natural preservatives to improve the quality of vacuum skin packaged beef during refrigerated storage. *Meat Sci*. 169: 108219.
130. Shaltout, F.A, Zakaria. I. M, Jehan Eltanani, Asmaa Elmelegy. (2015). Microbiological status of meat and chicken received to University student hostel. *BENHA VETERINARY MEDICAL JOURNAL*. 29(2): 187-192.
131. Selmi S, Rtibi K, Grami D, Sebai H, Marzouki L. (2017). Rosemary (*Rosmarinus officinalis*) essential oil components exhibit anti-hyperglycemic, anti-hyperlipidemic and antioxidant effects in experimental diabetes. *Pathophysiology*. 24(4): 297–303.
132. Saad,S.M.Edris, A.M. Shaltout,F.A. and Edris, Shima. (2012). Isolation and identification of salmonellae and E.coli from meat and poultry cuts by using A.multiplex PCR. *Benha Vet. Med.J*.16-26.

133. Arshad M.S, Amjad Z, Yasin M, Saeed F, Imran A, Sohaib M, Anjum F.M, Hussain S. (2019). Quality and stability evaluation of chicken meat treated with gamma irradiation and turmeric powder. *Int. J. Food Prop.* 22(1): 153–171.
134. Saad, S.M. and Shaltout, F.A. (1998). Mycological Evaluation of camel carcasses at Kalyobia Abattoirs. *Vet.Med.J. Giza.* 46(3): 223-229.
135. Smaoui S, Hlima H.B, Braïek O.B, Ennouri K, Mellouli L, Khaneghah A.M. (2021). Recent advancements in encapsulation of bioactive compounds as a promising technique for meat preservation. *Meat Sci.* 181:108585.
136. Beya M.M, Netzel M.E, Sultanbawa Y, Smyth H, Hoffman L.C. (2021). Plant-based phenolic molecules as natural preservatives in comminuted meats: A review. *Antioxidants.* 10(2):263.
137. Saad S.M, Hassanin, F. S, Shaltout, F.A, Marionette Z Nassif, Marwa Z Seif. (2019). Prevalence of Methicillin-Resistant *Staphylococcus Aureus* in Some Ready-to-Eat Meat Products. *American Journal of Biomedical Science & Research.* 4(6): 460-464.
138. Radunz M, Dos Santos Hackbart H.C, Camargo T.M, Nunes C.F.P, de Barros F.A.P, Dal Magro J, Filho P.J.S, Gandra E.A, Radunz A.L, da Rosa Zavareze E. (2020). Antimicrobial potential of spray drying encapsulated thyme (*Thymus vulgaris*) essential oil on the conservation of hamburger-like meat products. *Int. J. Food Microbiol.* 330: 108696.
139. Saad S.M, Shaltout, F.A, Nahla A Abou Elroos and Saber B El-nahas. (2019). Incidence of *Staphylococci* and *E. coli* in Meat and Some Meat Products. *EC Nutrition.* 14(6).
140. Zamuz S, Lopez-Pedrouso M, Barba F.J, Lorenzo J.M, Dominguez H, Franco D. (2018). Application of hull, bur and leaf chestnut extracts on the shelf-life of beef patties stored under MAP: Evaluation of their impact on physicochemical properties, lipid oxidation, antioxidant, and antimicrobial potential. *Food Res. Int.* 112: 263–273.
141. Shaltout FA, Riad EM, TES Ahmed and AbouElhassan A. (2017). Studying the Effect of Gamma Irradiation on Bovine Offal's Infected with *Mycobacterium tuberculosis* Bovine Type. *Journal of Food Biotechnology Research.* 1: 1-6.
142. Kiprotich S, Mendonca A, Dickson J, Shaw A, Thomas-Popo E, White S, Moutiq R, Ibrahim S.A. (2020). Thyme oil enhances the inactivation of *Salmonella enterica* on raw chicken breast meat during marination in lemon juice with added *Yucca schidigera* extract. *Front. Nutr.* 7: 619023.
143. Shaltout FA, Ahmed A A Maarouf and Mahmoud ES Elkhoully. (2017). Bacteriological Evaluation of Frozen Sausage. *Nutrition and Food Toxicology.* 1(5): 174-185.
144. Lourenço T, Mendonça E, Nalevaiko P, Melo R, Silva P, Rossi D. (2013). Antimicrobial effect of turmeric (*Curcuma longa*) on chicken breast meat contamination. *Braz. J. Poult. Sci.* 15(2): 79–82.
145. Shaltout FA, Zakaria IM and Nabil ME. (2018). Incidence of Some Anaerobic Bacteria Isolated from Chicken Meat Products with Special Reference to *Clostridium perfringens*. *Nutrition and Food Toxicology.* 2(5): 429-438.
146. Lee N.-K, Jung B.S, Na D.S, Yu H.H, Kim J.-S, Paik H.-D. (2016). The impact of antimicrobial effect of chestnut inner shell extracts against *Campylobacter jejuni* in chicken meat. *LWT.* 65: 746–750.
147. Shaltout FA, Mohamed, A.Hassan and Hassanin, F. S. (2004). THERMAL INACTIVATION OF ENTEROHAEMORRHAGIC *ESCHERICHIA COLI* O157:H7 AND ITS SENSITIVITY TO NISIN AND LACTIC ACID CULTURES.1: 336-334.
148. Hulankova R, Borilova G, Steinhauserova I. (2013). Combined antimicrobial effect of oregano essential oil and caprylic acid in minced beef. *Meat Sci.* 95(2): 190–194.
149. Shaltout FA, El-diahy, E.M. Elmesalamy, M. and Elshaer, M. (2014). Study on fungal contamination of some chicken meat products with special reference to the use of PCR for its identification. Conference, *Veterinary Medical Journal – Giza.* 60: 1-10.
150. Hernandez H, Frankova A, Sykora T, Kloucek P, Kourimska L, Kucerova I, Banout J. (2017). The effect of oregano essential oil on microbial load and sensory attributes of dried meat. *J. Sci. Food Agric.* 97(1): 82–87.
151. Shaltout, F.A. (2002). Microbiological Aspects of Semi-cooked chicken Meat Products. *Benha Veterinary Medical Journal.* 13(2): 15-26.
152. Shange N, Makasi T, Gouws P, Hoffman L.C. (2019). Preservation of previously frozen black wildebeest meat (*Connochaetes gnou*) using oregano (*Oreganum vulgare*) essential oil. *Meat Sci.* 148: 88–95.
153. Lages L.Z, Radünz M, Gonçalves B.T, Silva da Rosa R, Fouchy M.V, de Cássia dos Santos da Conceição R, Gularte M.A, Barboza Mendonça C.R, Gandra E.A. (2021). Microbiological and sensory evaluation of meat sausage using thyme (*Thymus vulgaris*, L.) essential oil and powdered beet juice (*Beta vulgaris* L, Early Wonder cultivar) *LWT.* 148: 109896.
154. Shaltout FA, Mohammed Farouk; Hosam A.A. Ibrahim and Mostafa E.M. Afifi. (2017). Incidence of Coliform and *Staphylococcus aureus* in ready to eat fast foods. *BENHA VETERINARY MEDICAL JOURNAL.* 32(1): 13 - 17.
155. De Oliveira T.L, de Araujo Soares R, Ramos E.M, das Gracias Cardoso M, Alves E, Piccoli R.H. (2011). Antimicrobial activity of *Satureja montana* L. essential oil against *Clostridium perfringens* type A inoculated in mortadella-type sausages formulated with different levels of sodium nitrite. *Int. J. Food Microbiol.* 144(3): 546–555.
156. Shaltout, F.A, Zakaria, I.M, Nabil, M.E. (2017). Detection and typing of *Clostridium perfringens* in some retail chicken meat products. *BENHA VETERINARY MEDICAL JOURNAL.* 33(2): 283-291.

157. Zhu Y, Li C, Cui H, Lin L. (2020). Encapsulation strategies to enhance the antibacterial properties of essential oils in food system. *Food Control*. 123(2): 107856.
158. Shaltout, F.A. (1992). Studies on Mycotoxins in Meat and Meat by Products. M.V.Sc Thesis Faculty of Veterinary Medicine, Moshtohor, Zagazig University Benha branch.
159. Juneja V.K, Fan X, Peña-Ramos A, Diaz-Cinco M, Pacheco-Aguilar R. (2006). The effect of grapefruit extract and temperature abuse on growth of *Clostridium perfringens* from spore inocula in marinated, sous-vide chicken products. *Innov. Food Sci. Emerg. Technol.* 7(1): 100–106.
160. Shaltout, F.A. (1996). Mycological And Mycotoxicological profile Of Some Meat products. Ph.D.Thesis, Faculty of Veterinary Medicine, Moshtohor, Zagazig University Benha branch.
161. Maes C, Bouquillon S, Fauconnier M.-L. (2019). Encapsulation of essential oils for the development of biosourced pesticides with controlled release: A review. *Molecules*. 24(14): 2539.
162. Shaltout, F.A. (1998). Proteolytic Psychrotrophes in Some Meat products. *Alex. Vet. Med. J.* 14(2): 97-107.
163. Ben-Fadhel Y, Cingolani M.C, Li L, Chazot G, Salmieri S, Horak C, Lacroix M. (2021). Effect of γ -irradiation and the use of combined treatments with edible bioactive coating on carrot preservation. *Food Packag. Shelf Life*. 28(1): 100635.
164. Shaltout, F.A. (1999). Anaerobic Bacteria in Vacuum Packed Meat Products. *Benha Vet. Med.J.* 10(1): 1-10.
165. Soyer F, Keman D, Eroglu E, Ture H. (2020). Synergistic antimicrobial effects of activated lactoferrin and rosemary extract in vitro and potential application in meat storage. *J. Food Sci. Technol.* 57(12): 4395–4403.
166. Shaltout, F.A. (2000). Protozoal Foodborne Pathogens in some Meat Products. *Assiut Vet. Med. J.* 42(84): 54-59.
167. Kahraman T, Issa G, Bingol E.B, Kahraman B.B, Dumen E. (2015). Effect of rosemary essential oil and modified-atmosphere packaging (MAP) on meat quality and survival of pathogens in poultry fillets. *Braz. J. Microbiol.* 46(2): 591–599.
168. Shaltout, F.A. (2001). Quality evaluation of sheep carcasses slaughtered at Kalyobia abattoirs. *Assiut Veterinary Medical Journal*. 46(91): 150-159.
169. Motavaf F, Mirvaghefi A, Farahmand H, Hosseini S.V. (2021). Effect of *Zataria multiflora* essential oil and potassium sorbate on inoculated *Listeria monocytogenes*, microbial and chemical quality of raw trout fillet during refrigerator storage. *Food Sci. Nutr.* 9(6): 3015–3025.
170. Shaltout, F.A. (2002). Microbiological Aspects of Semi-cooked Chicken Meat Products. *Benha Vet. Med.J.* 13(2): 15-26.
171. Sojic B, Pavlic B, Ikonc P, Tomovic V, Ikonc B, Zekovic Z, Kocic-Tanackov S, Jokanovic M, Skaljic S, Ivic M. (2019). Coriander essential oil as natural food additive improves quality and safety of cooked pork sausages with different nitrite levels. *Meat Sci.* 157: 107879.
172. Shaltout, F.A. (2003). *Yersinia Enterocolitica* in some meat products and fish marketed at Benha city. The Third international conference Mansoura 29-30 April. 1.
173. Asioli D, Aschemann-Witzel J, Caputo V, Vecchio R, Annunziata A, Næs T, Varela P. (2017). Making sense of the “clean label” trends: A review of consumer food choice behavior and discussion of industry implications. *Food Res. Int.* 99: 58–71.
174. Shaltout, F.A. (2009). Microbiological quality of chicken carcasses at modern Poultry plant. The 3rd Scientific Conference, Faculty of Vet. Med, Benha University. 137-150.
175. Coutinho de Oliveira T.L, Malfitano de Carvalho S, de Araújo Soares R., Andrade M.A, Cardoso M.d.G, Ramos E.M, Piccoli R.H. (2012). Antioxidant effects of *Satureja montana* L. essential oil on TBARS and color of mortadella-type sausages formulated with different levels of sodium nitrite. *LWT*. 45: 204–212.
176. Shaltout, F.A. and Abdel Aziz, A.M. (2004). *Salmonella enterica* Serovar Enteritidis in Poultry Meat and their Epidemiology. *Vet. Med. J., Giza*. 52(3): 429-436.
177. World Health Organization. High-Dose Irradiation: Wholesomeness of Food Irradiated with Doses above 10 kGy. (1997). World Health Organization; Geneva, Switzerland.
178. Shaltout, F.A. and Abdel Aziz, A.M. (2004). *ESCHERICHIA COLI* STRAINS IN SLAUGHTERED ANIMALS AND THEIR PUBLIC HEALTH IMPORTANCE. *J. Egypt. Vet. Med. Association*. 64(2): 7-21.
179. Abdeldaiem M. (2014). Using of combined treatment between edible coatings containing ethanolic extract of papaya (*Carica papaya* L.) leaves and gamma irradiation for extending shelf-life of minced chicken meat. *Am. J. Food Technol.* 2(1): 6–16.
180. Shaltout, F.A, Amin, R, Marionet, Z, Nassif and Shimaa, Abdel-wahab. (2014). Detection of aflatoxins in some meat products. *Benha veterinary medical journal*. 27(2): 368-374.
181. Akhter R, Masoodi F, Wani T.A, Rather S.A, Hussain P.R. (2021). Synergistic effect of low dose γ -irradiation, natural antimicrobial and antioxidant agents on quality of meat emulsions. *Radiat. Phys. Chem.* 189(1): 109724.
182. Shaltout, F.A. and Afify, Jehan Riad, EM and Abo Elhasan, Asmaa, A. (2012). Improvement of microbiological status of oriental sausage. *Journal of Egyptian Veterinary Medical Association*. 72(2): 157-167.
183. European Commission EU Guidance to the Commission Regulation (EC) No 450/2009 of 29 May 2009 on Active and Intelligent Materials and Articles Intended to Come into the Contact with Food. [(accessed on 12 October 2021)]. Available online.
184. Shaltout, F.A. and Daoud, J. R. (1996). Chemical analytical studies on rabbit meat and liver. *Benha Vet. Med.J.* 7(2): 55-64.
185. Ming Y, Chen L, Khan A, Wang H, Wang C. (2020). Effects of tea polyphenols on physicochemical and antioxidative properties of whey protein coating. *Food Sci. Biotechnol.* 29(12): 1655–1663.

186. Shaltout, F.A. and Edris, A.M. (1999). Contamination of shawerma with pathogenic yeasts. *Assiut Veterinary Medical Journal*. 40(64): 34-39.
187. Balasubramaniam V, Martinez-Monteagudo S.I, Gupta R. (2015). Principles and application of high pressure-based technologies in the food industry. *Annu. Rev. Food Sci.* 6: 435–462.
188. Shaltout, F. A. Eldiasty, E. and Mohamed, M.S. (2014). Incidence of lipolytic and proteolytic fungi in some chicken meat products and their public health significance. *Animal Health Research Institute: First International Conference on Food Safety and Technology 19-23 June 2014 Cairo Egypt*. 79-89.
189. Chuang S, Sheen S. (2021). High pressure processing of raw meat with essential oils-microbial survival, meat quality, and models: A review. *Food Control*. 132(35): 108529.
190. Shaltout, F.A.Eldiasty, E. Salem, R. and Hassan, Asmaa. (2016). Mycological quality of chicken carcasses and extending shelf – life by using preservatives at refrigerated storage. *Veterinary Medical Journal -Giza (VMJG)*. 62(3): 1-7.
191. Pedreschi F, Mariotti-Celis M.S. *Genetically Modified and Irradiated Food*. Academic Press; Cambridge, MA, USA 2020. Irradiation kills microbes: Can it do anything harmful to the food. 233–242.
192. Shaltout, F.A.Salem, R. Eldiasty, E. and Diab, Fatema. (2016). Mycological evaluation of some ready to eat meat products with special reference to molecular characterization. *Veterinary Medical Journal -Giza*. 62(3): 9-14.
193. Global Newswire The “Clean Label Ingredient Market–Growth, Trends, and Forecast (2018–2023)”. [(accessed on 12 October 2021)]. Available online.
194. Shaltout, F. A. Elshater, M. and Wafaa, Abdelaziz. (2015). Bacteriological assessment of street vended meat products sandwiches in Kalyobia Governorate. *Benha Vet. Med.J.* 28(2): 58-66.
195. Luong N.-D.M, Coroller L, Zagorec M, Membré J.-M, Guillou S. (2020). Spoilage of chilled fresh meat products during storage: A quantitative analysis of literature data. *Microorganisms*. 8(8): 1198.
196. Shaltout, F. A. Gerges, M.T. and Shewail, A.A. (2018). Impact of Organic Acids and Their Salts on Microbial Quality and Shelf Life of Beef. *Assiut veterinary medical journal*. 64(159): 164-177.
197. Martillanes S, Rocha-Pimienta J, Llera-Oyola J, Gil M.V, Ayuso-Yuste M.C, García-Parra J, Delgado-Adámez J. (2021). Control of *Listeria monocytogenes* in sliced dry-cured Iberian ham by high pressure processing in combination with an eco-friendly packaging based on chitosan, nisin and phytochemicals from rice bran. *Food Control*. 124: 107933.
198. Shaltout, F.A.Ghoneim, A.M. Essmail, M.E. and Yousseif ,A. (2001). Studies on aflatoxin B1 residues in rabbits and their pathological effects. *J.Egypt. Vet. Med. Association*. 61(2): 85-103.
199. Lee J.-S, Choi Y.S, Lee H.G. (2020). Synergistic antimicrobial properties of nanoencapsulated clove oil and thymol against oral bacteria. *Food Sci. Biotechnol.* 29(11): 1597–1604.
200. Shaltout, F.A. and Hanan, M.T. El-Lawendy. (2003). Heavy Metal Residues In Shawerma. *Beni-Suef Vet.Med.J.* 13(1): 213-224.
201. Park S, Mun S, Kim Y.-R. (2020). Influences of added surfactants on the water solubility and antibacterial activity of rosemary extract. *Food Sci. Biotechnol.* 29(10): 1373–1380.
202. Shaltout, F.A. and Hashim, M.F. (2002). Histamine in salted, Smoked and Canned Fish products. *Benha Vet. Med.J.* 13(1): 1-11.
203. Fang Z, Zhao Y, Warner R.D, Johnson S.K. (2017). Active and intelligent packaging in meat industry. *Trends Food Sci. Technol.* 61: 60–71.
204. Shaltout, F.A. Hashim, M.F. and Elnahas, s. (2015). Levels of some heavy metals in fish (*tilapia nilotica* and *Claris lazera*) at Menufia Governorate. *Benha Vet. Med.J.* 29(1): 56-64.
205. Yong H.I, Kim T.K, Choi H.D, Jang H.W, Jung S, Choi Y.S. (2021). Clean label meat technology: Pre-converted nitrite as a natural curing. *Food Sci. Anim. Resour.* 41(2): 173–184.
206. Shaltout, F.A. and Ibrahim, H.M. (1997). Quality evaluation of luncheon and Alexandrian sausage. *Benha Vet. Med.J.* 10(1): 1-10.
207. Barcenilla C, Ducic M, López M, Prieto M, Álvarez-Ordóñez A. (2022). Application of lactic acid bacteria for the biopreservation of meat products: A systematic review. *Meat Sci.* 183: 108661.
208. Shaltout, F.A. Nassif, M and Shakran, A. (2014). Quality of battered and breaded chicken meat products. *Global Journal of Agriculture and Food Safety Science*. 1(2): 283-299.
209. Crowe W, Elliott C.T, Green B.D. (2019). A review of the in vivo evidence investigating the role of nitrite exposure from processed meat consumption in the development of colorectal cancer. *Nutrients*. 11(11): 2673.
210. Shaltout, F.A, Amani M. Salem, A. H. Mahmoud, K. A. (2013). Bacterial aspect of cooked meat and offal at street vendors level. *Benha veterinary medical journal*. 24(1): 320-328.
211. Choe E. (2020). Roles and action mechanisms of herbs added to the emulsion on its lipid oxidation. *Food Sci. Biotechnol.* 29(9):1165–1179.
212. Shaltout, F.A. and Salem, R.M. (2000). Moulds, aflatoxin B1 and Ochratoxin A in Frozen Livers and meat products. *Vet . Med. J.Giza*. 48(3): 341-346.
213. Marrone R, Smaldone G, Ambrosio R.L, Festa R, Ceruso M, Chianese A, Anastasio A. (2021). Effect of beetroot (*Beta vulgaris*) extract on black angus burgers shelf life. *Ital. J. Food Saf.* 10(1): 9031.
214. Yasser H. Al-Tarazi, A. Al-Zamil, Shaltout FA. and H. Abdel-Samei. (2002). Microbiological status of raw cow milk marketed in northern Jordan. *AVMJ*. 49(96): 180-194.
215. Lee N.-K, Paik H.-D. (2016). Status, Antimicrobial mechanism, and regulation of natural preservatives in livestock food systems. *Korean J. Food Sci. Anim. Resour.* 36(4): 547–557.
216. Shaltout FA, Zakaria IM and Nabil ME. (2018). Incidence of Some Anaerobic Bacteria Isolated from Chicken Meat Products with

- Special Reference to *Clostridium perfringens*. *Nutrition and Food Toxicology*. 2(5): 429-438.
217. Olszewska M.A, Gędas A, Simões M. (2020). Antimicrobial polyphenol-rich extracts: Applications and limitations in the food industry. *Food Res. Int.* 134: 109214.
218. Shaltout, F. A. El-diasty, E.M. and Mohamed, M. S. (2014). Incidence of lipolytic and proteolytic fungi in some chicken meat products and their public health significance. 1st Scientific conference of food safety and Technology. 79-89.
219. Chaleshtori F.S, Arian A, Chaleshtori R.S. (2018). Assessment of sodium benzoate and potassium sorbate preservatives in some products in Kashan, Iran with estimation of human health risk. *Food Chem. Toxicol.* 120: 634-638.
220. Shaltout, F. A. El-diasty, E.M. Salem, R. M. and Asmaa, M. A. Hassan. (2016). Mycological quality of chicken carcasses and extending shelf -life by using preservatives at refrigerated storage. *Veterinary Medical Journal – Giza*. 62(3) :1-10.
221. International Agency for Research on Cancer (IARC) monographs on the evaluation of carcinogenic risks to humans Ingested nitrate and nitrite, and cyanobacterial peptide toxins. (2010). *IARC Monogr. Eval. Carcinog. Risks Hum.* 1-412.
222. Shaltout FA, R.M. Salem, E.M. El-Diasty and W.I.M. Hassan. (2019). Effect of Lemon Fruits and Turmeric Extracts on Fungal Pathogens in Refrigerated Chicken Fillet Meat. *Global Veterinaria*. 21(3): 156-160.
223. Cao Q, Yan J, Sun Z, Gong L, Wu H, Tan S, Lei Y, Jiang B, Wang Y. (2021). Simultaneous optimization of ultrasound-assisted extraction for total flavonoid content and antioxidant activity of the tender stem of *Triarrhena lutarioriparia* using response surface methodology. *Food Sci. Biotechnol.* 30(1): 37–45.
224. Shaltout FA, El-diasty, E, M. Elmesalamy, M. and Elshaer, M. (2014). Study on fungal contamination of some chicken meat products with special reference to 2 the use of PCR for its identification. Conference, *Veterinary Medical Journal – Giza*. 60: 1-10.
225. Piper J.D, Piper P.W. (2017). Benzoate and sorbate salts: A systematic review of the potential hazards of these invaluable preservatives and the expanding spectrum of clinical uses for sodium benzoate. *Compr. Rev. Food Sci. Food Saf.* 16(5): 868–880.
226. Shaltout, F. A. Salem, R. M El-diasty, Eman and Fatema, A.H. Diab. (2016). Mycological evaluation of some ready to eat meat products with special reference to molecular characterization. *Veterinary Medical Journal – Giza*. 62(3): 9-14.
227. Shim S.-M, Seo S.H, Lee Y, Moon G.-I, Kim M.-S, Park J.-H. (2011). Consumers' knowledge and safety perceptions of food additives: Evaluation on the effectiveness of transmitting information on preservatives. *Food Control*. 22(7): 1054–1060.
228. Shaltout FA, Ahmed, A.A. Maarouf, Eman, M.K. Ahmed. (2018). Heavy Metal Residues in chicken cuts up and processed chicken meat products. *BENHA VETERINARY MEDICAL JOURNAL*. 34(1): 473-483.
229. Ministry of Food and Drug Safety (MFDS) Food Additives Code. [(accessed on 12 October 2021)]. Available online.
230. Shaltout, F.A. Hanan M. Lamada, Ehsan A.M. Edris. (2020). Bacteriological examination of some ready to eat meat and chicken meals. *Biomed J Sci & Tech Res*. 27(1): 20461- 20465.
231. Matthews K.R, Kniel K.E, Montville T.J. (2017). *Food Microbiology: An Introduction*. 4th ed. ASM Press; Washington, DC, USA.
232. Sobhy, Asmaa and Shaltout, Fahim. (2020). Prevalence of some food poisoning bacteria in semi cooked chicken meat products at Qaliubiya governorate by recent Vitek 2 compact and PCR techniques. *Benha Veterinary Medical Journal*. 38(2): 88-92.
233. Bohrer B.M. (2017). Nutrient density and nutritional value of meat products and non-meat foods high in protein. *Trends Food Sci. Technol.* 65: 103–112.
234. Sobhy, Asmaa and Shaltout, Fahim. (2020). Detection of food poisoning bacteria in some semi-cooked chicken meat products marketed at Qaliubiya governorate. *Benha Veterinary Medical Journal*. 38(2): 93-96.
235. Zhou G, Xu X, Liu Y. (2010). Preservation technologies for fresh meat—A review. *Meat Sci*. 86(1): 119–128.
236. Shaltout, F.A. (2024). Abattoir And Bovine Tuberculosis as A Reemerging Foodborne Diseases. *Clinical Medical Reviews and Report*. 6(1): 1-7.
237. World Health Organization. (2015). WHO Estimates of the Global Burden of Foodborne Diseases: Foodborne Disease Burden Epidemiology Reference Group 2007–2015. World Health Organization (WHO); Geneva, Switzerland: 1–15.
238. Shaltout, F.A. (2023). Viruses in Beef, Mutton, Chevron, Venison, Fish and Poultry Meat Products. *Food Science & Nutrition Technology*. 8(4): 1-10.
239. Lee H, Yoon Y. (2021). Etiological agents implicated in foodborne illness worldwide. *Food Sci. Anim. Resour*. 41(1): 1–7.
240. Farnaud S, Evans R.W. (2003). Lactoferrin—A multifunctional protein with antimicrobial properties. *Mol. Immunol*. 40(7): 395-405.